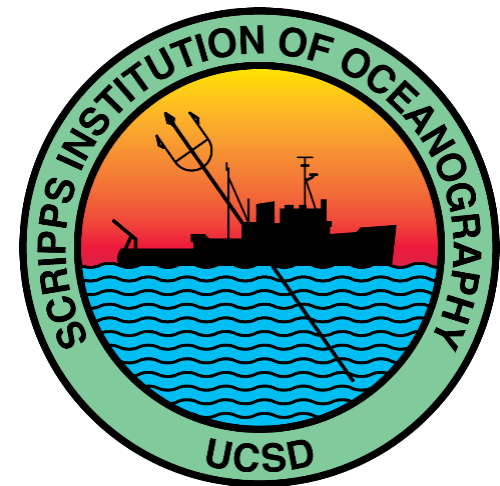


Optics of Marine Particles

Lecture 2

Dariusz Stramski

Scripps Institution of Oceanography
University of California San Diego
Email: dstramski@ucsd.edu



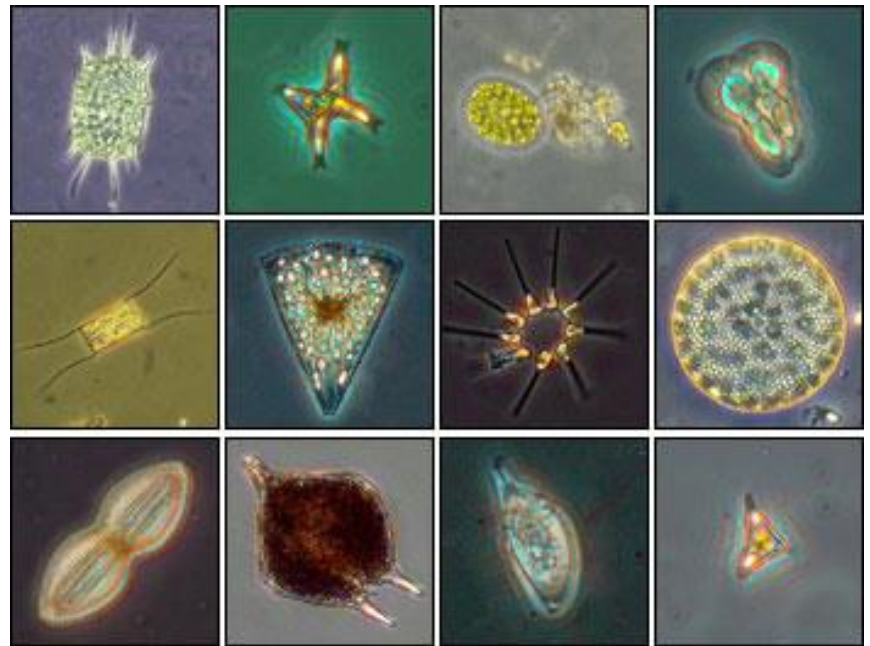
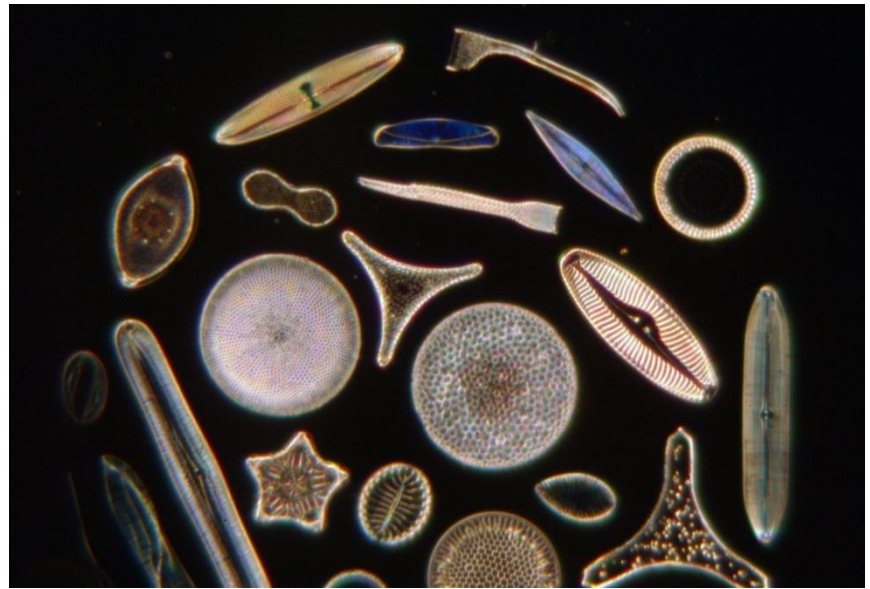
IOCCG Summer Lecture Series

22 July - 2 August 2014, Villefranche-sur-Mer, France

Suspended Particulate Matter

- Plankton microorganisms
- Biogenic detrital particles
- Mineral particles
- Colloidal particles
- Air bubbles

Plankton microorganisms

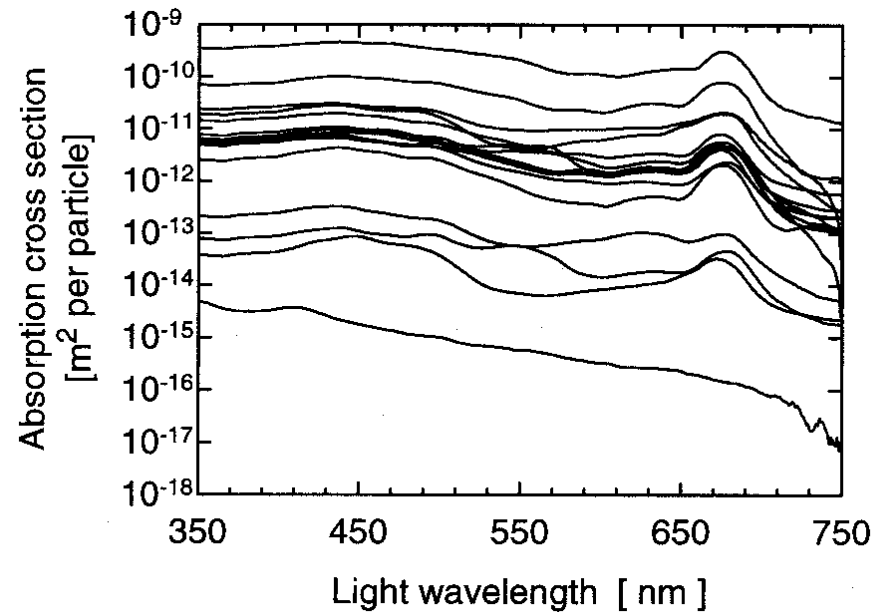
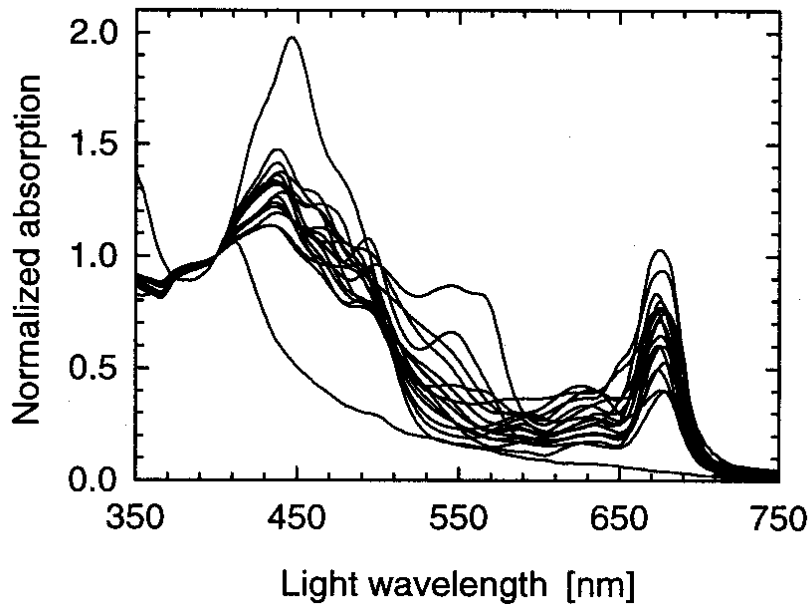
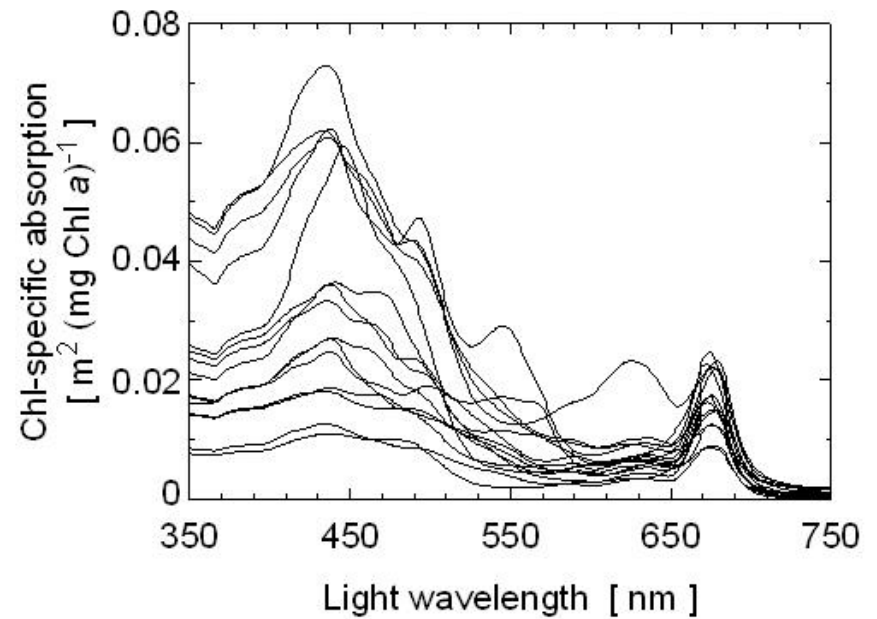


Plankton microorganisms

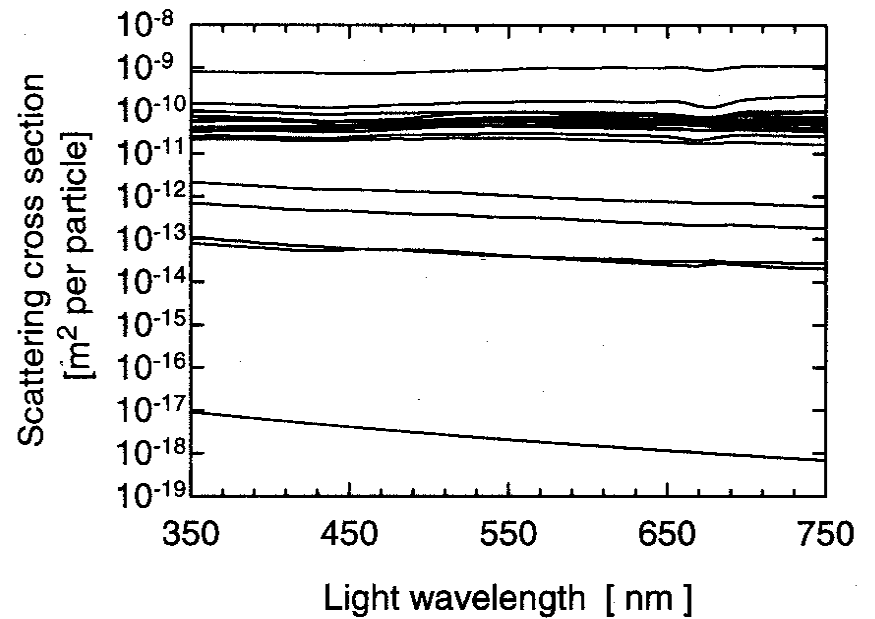
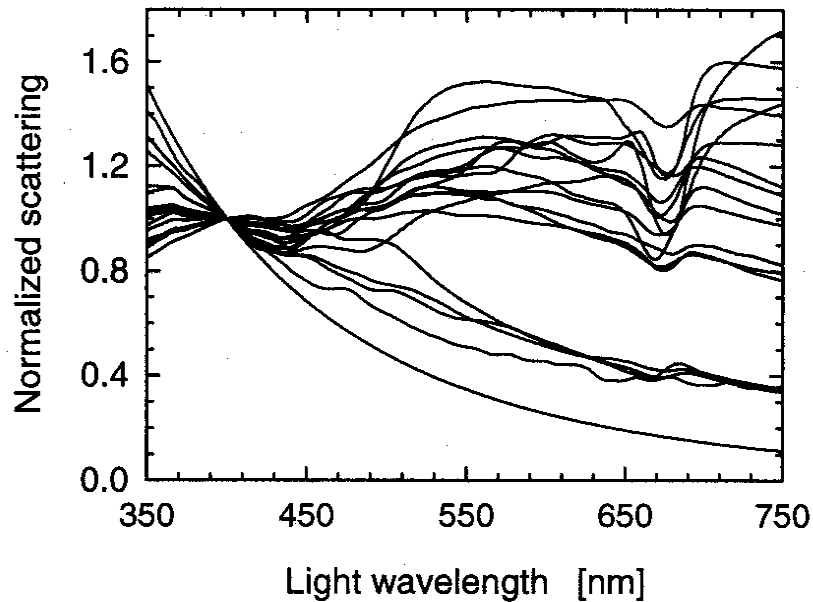
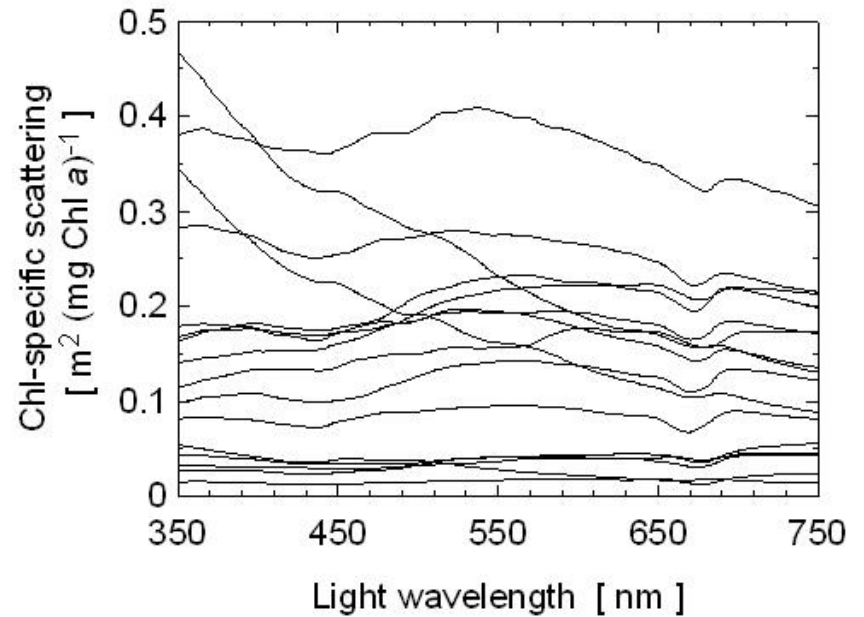
Table 1. Microbial components in the database and source of raw data. Values for the average equivalent spherical diameter (D), the real part of refractive index at 550 nm (n), and imaginary part of refractive index at 440 and 675 nm (n') are also given for each component.

i	Label	Microbial species	D [μm]	n 550 nm	n' · 10 ³ 440 nm	n' · 10 ³ 675 nm	Source of raw data
1	VIRU	Viruses	0.07	1.050	0	0	Stramski and Kiefer, 1991
2	HBAC	Heterotrophic bacteria	0.55	1.055	0.509	0.057	Stramski and Kiefer, 1990
3	PROC	generic Prochlorophyte; the average of:	0.66	1.051	18.51	10.30	
		PMED - Prochlorococcus strain MED	0.59	1.055	23.25	13.77	Morel et al., 1993
		PNAS - average of Prochlorococcus strains NATL and SARG	0.70	1.046	13.78	6.687	Morel et al., 1993
4	SYNE	generic Synechococcus; the average of:	1.05	1.051	5.587	2.930	
		SM41 - Synechococcus strain MAX41 (Cyanophyceae)	0.92	1.047	5.415	2.905	Morel et al., 1993
		SM01 - Synechococcus strain MAX01 (Cyanophyceae)	0.94	1.049	4.505	2.547	Morel et al., 1993
		SROS - Synechococcus strain ROS04 (Cyanophyceae)	1.08	1.049	4.516	2.154	Morel et al., 1993
		SDC2 - Synechococcus strain DC2 (Cyanophyceae)	1.14	1.050	4.249	2.375	Morel et al., 1993
		SI03 - Synechococcus strain WH8103 (Cyanophyceae)	1.14	1.062	9.251	4.668	Stramski et al., 1995
5	SYMA	generic phycocyanin-rich picophytoplankton; the average of:	1.41	1.055	6.495	2.757	
		SCYS - Synechocystis (Cyanophyceae)	1.39	1.050	4.530	1.910	Ahn et al., 1992
		MARI - Anacystis marina (Cyanophyceae)	1.43	1.060	8.460	3.603	Ahn et al., 1992
6	PING	Pavlova pinguis (Haptophyceae)	3.97	1.046	4.177	2.709	Bricaud et al., 1988
7	PSEU	Thalassiosira pseudonana (Bacillariophyceae)	3.99	1.045	9.231	7.397	Stramski and Reynolds, 1993
8	LUTH	Pavlova lutheri (Haptophyceae)	4.26	1.045	5.767	2.403	Bricaud et al., 1988
9	GALB	Isochrysis galbana (Haptophyceae)	4.45	1.056	7.673	5.101	Ahn et al., 1992
10	HUXL	Emiliana huxleyi (Haptophyceae)	4.93	1.050	5.012	2.950	Ahn et al., 1992
11	CRUE	Porphyridium cruentum (Rhodophyceae)	5.22	1.051	3.351	2.443	Bricaud et al., 1988
12	FRAG	Chroomonas fragarioides (Cryptophyceae)	5.57	1.039	4.275	2.904	Ahn et al., 1993
13	PARV	Prymnesium parvum (Haptophyceae)	6.41	1.045	2.158	1.329	Bricaud et al., 1988
14	BIOC	Dunaliella bioculata (Chlorophyceae)	6.71	1.038	10.49	7.839	Ahn et al., 1993
15	TERT	Dunaliella tertiolecta (Chlorophyceae)	7.59	1.063	6.260	5.076	Stramski et al., 1993
16	CURV	Chaetoceros curvisetum (Bacillariophyceae)	7.73	1.024	2.877	1.480	Bricaud et al., 1988
17	ELON	Hymenomonas elongata (Haptophyceae)	11.77	1.046	13.87	7.591	Ahn et al., 1992
18	MICA	Prorocentrum micans (Dinophyceae)	27.64	1.045	2.466	1.710	Ahn et al., 1992

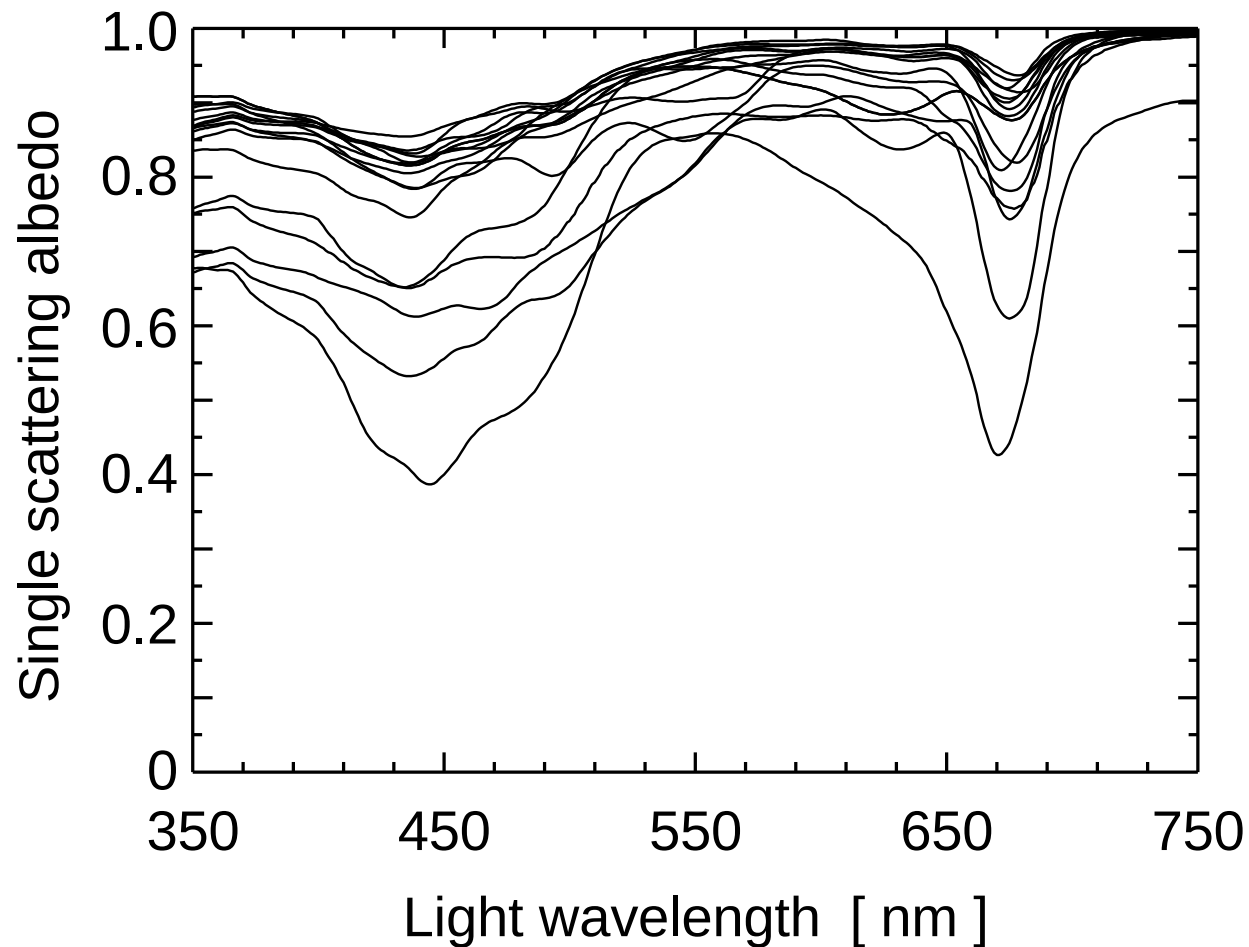
Interspecies variability in absorption



Interspecies variability in scattering

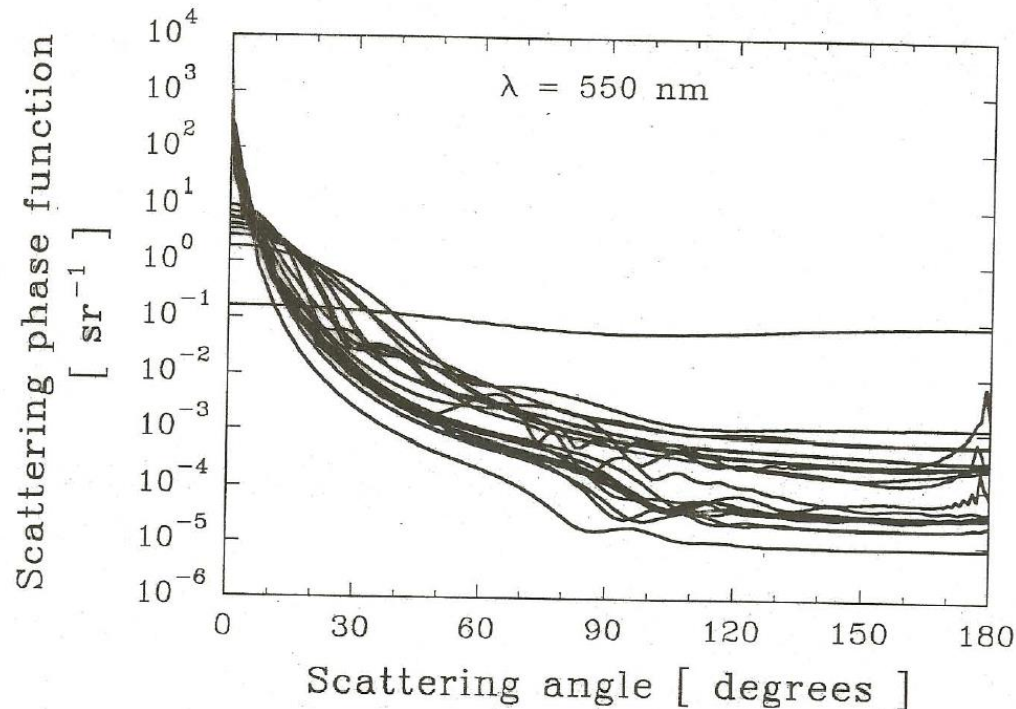


Interspecies variability in single scattering albedo



(based on data from Stramski et al. 2001)

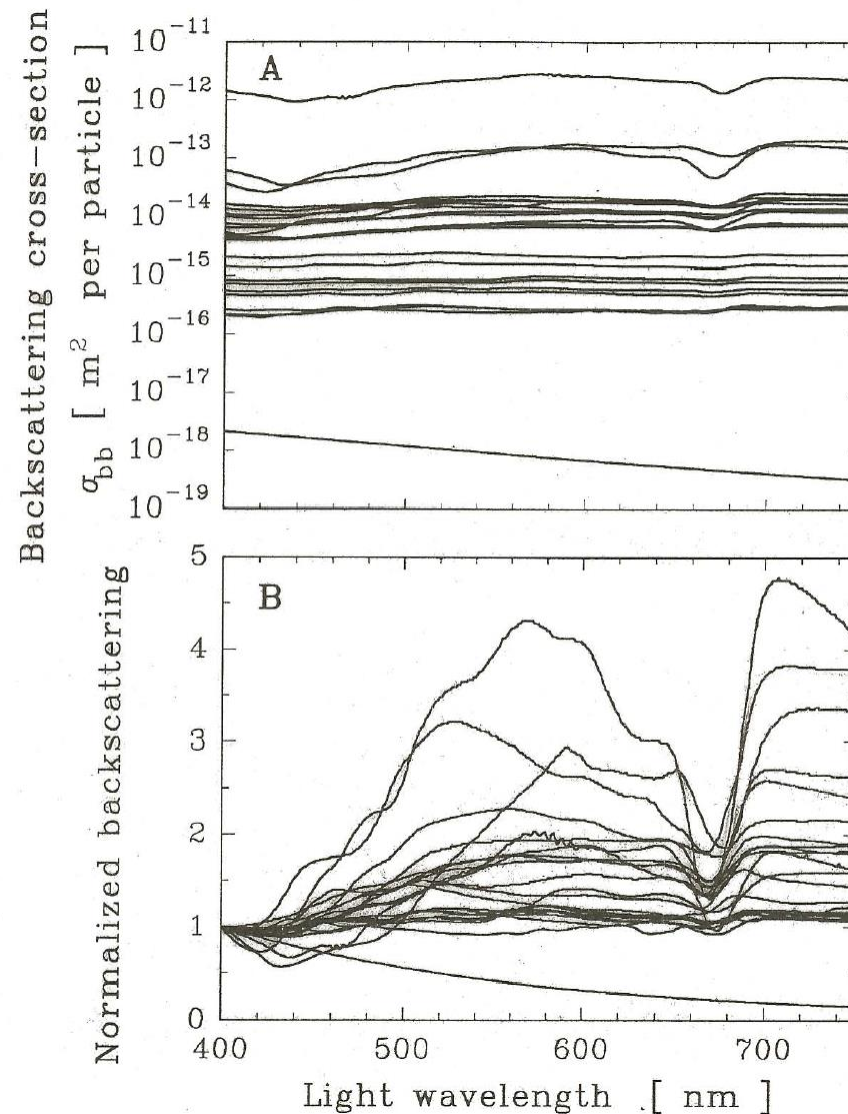
Mie calculations of scattering phase function for plankton microorganisms



Viruses
Heterotrophic bacteria
Prochlorococcus (2 strains)
Synechococcus (Cyanophyceae, 5 strains)
Anacystis marina (Cyanophyceae)
Pavlova pinguis (Haptophyceae)
Thalassiosira pseudonana (Bacillariophyceae)
Pavlova lutheri (Haptophyceae)
Isochrysis galbana (Haptophyceae)
Emiliana huxleyi (Haptophyceae)
Porphyridium cruentum (Rhodophyceae)
Chroomonas fragarioides (Cryptophyceae)
Prymnesium parvum (Haptophyceae)
Dunaliella bioculata (Chlorophyceae)
Dunaliella tertiolecta (Chlorophyceae)
Chaetoceros curvisetum (Bacillariophyceae)
Hymenomonas elongata (Haptophyceae)
Prorocentrum micans (Dinophyceae)

Backscattering properties of plankton microorganisms

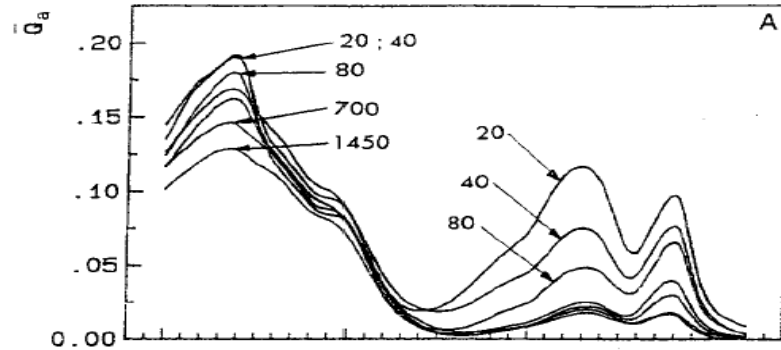
(subject to uncertainties associated with Mie scattering calculations for homogeneous spheres)



(Stramski et al. 2001)

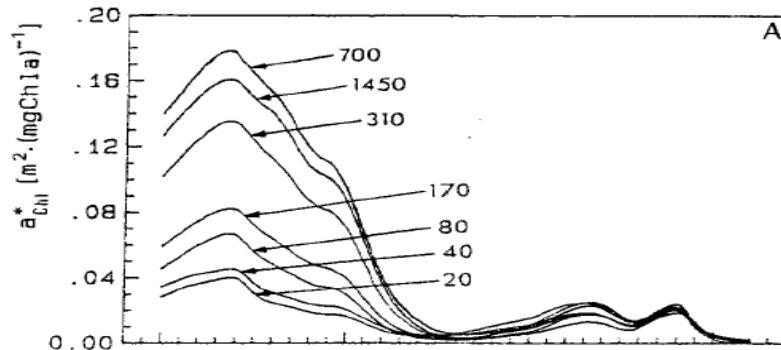
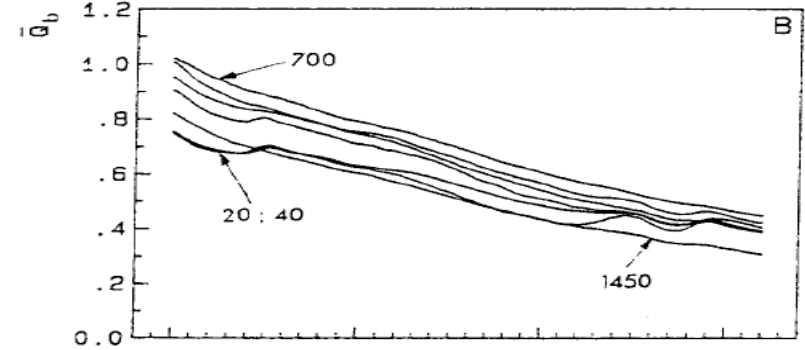
Intraspecies variability due to irradiance - *Synechocystis*

Absorption

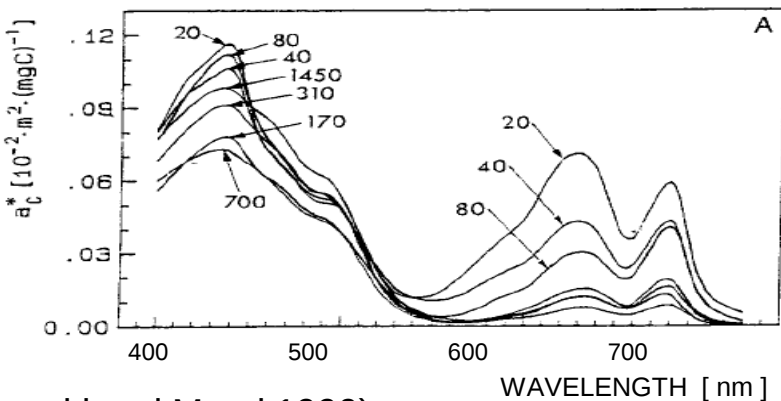
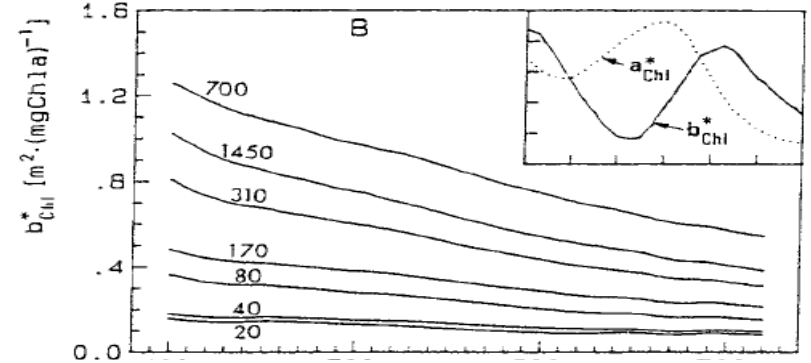


Cell

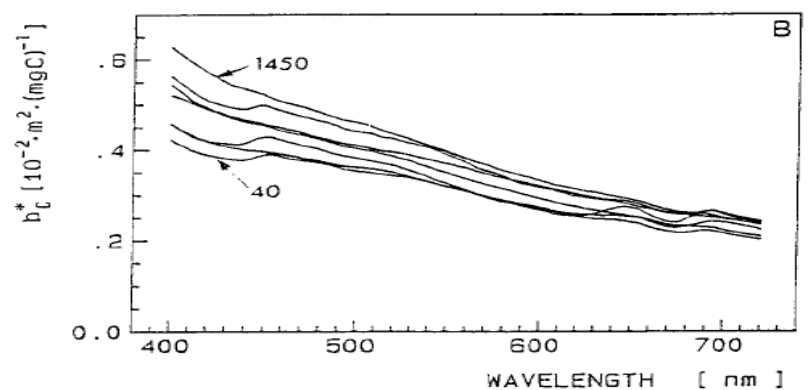
Scattering



Chl

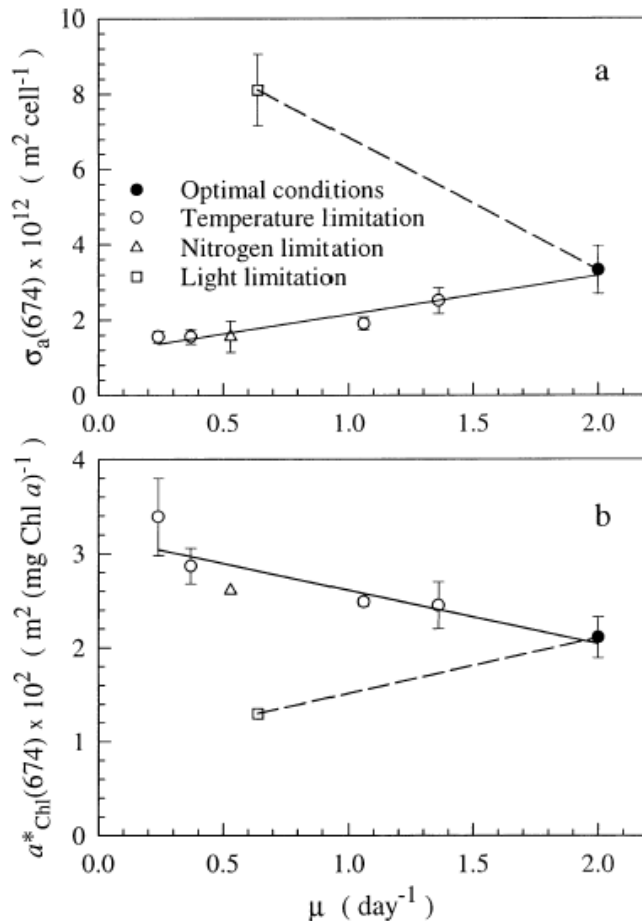


C

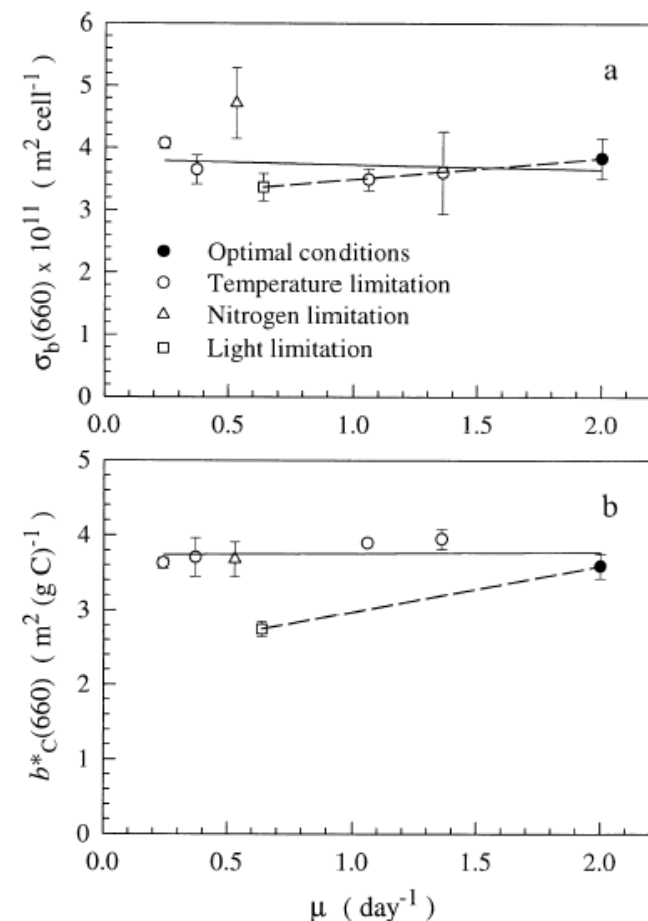


Intraspecies variability due to temperature, nitrogen, and light limitation – *Thalassiosira pseudonana*

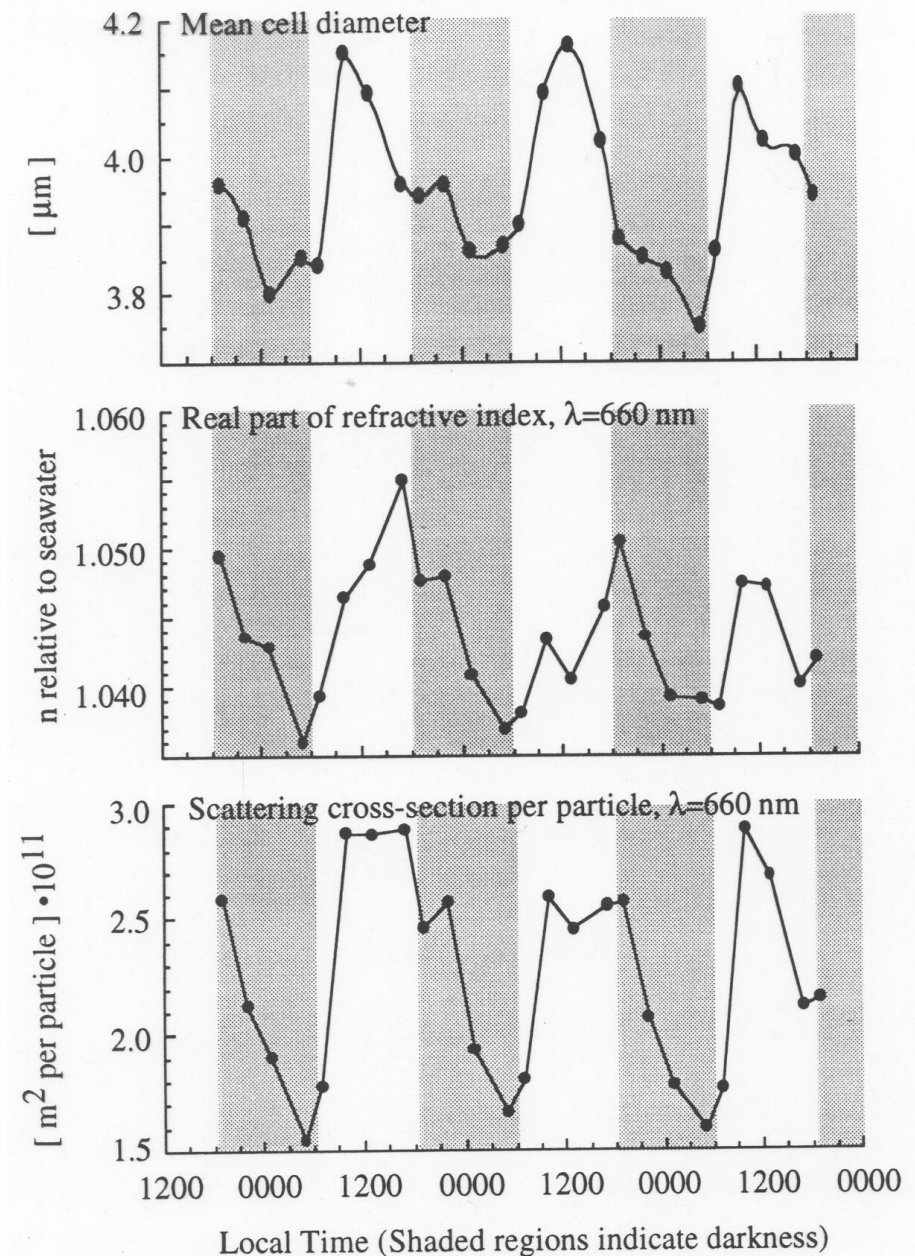
Absorption vs. growth rate



Scattering vs. growth rate



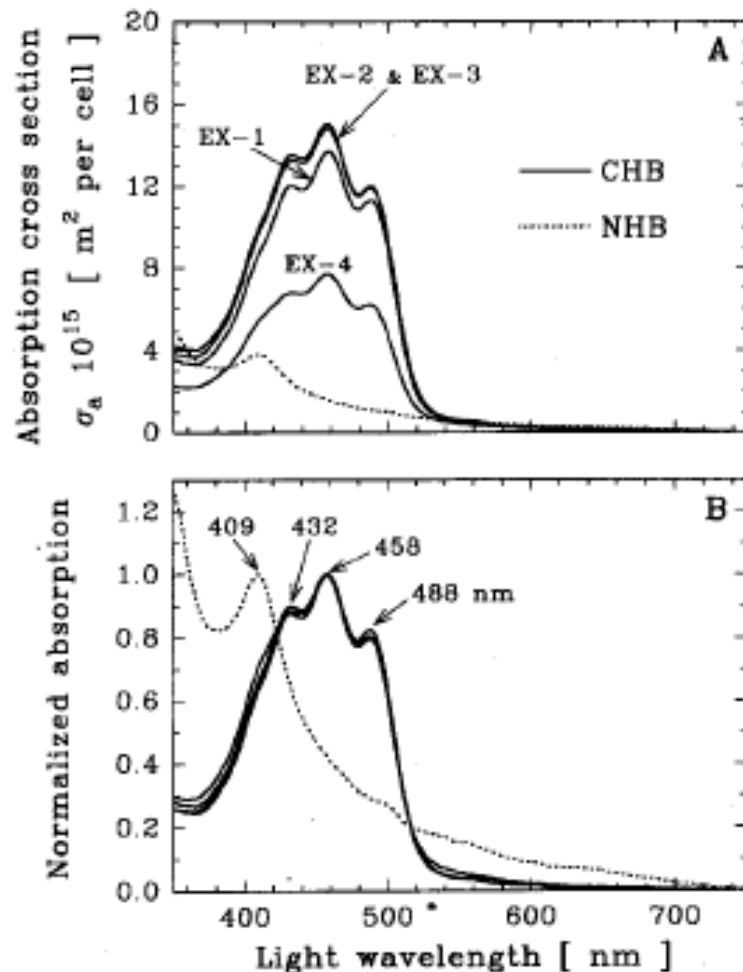
Intraspecies
variability over
a diel cycle
*Thalassiosira
pseudonana*



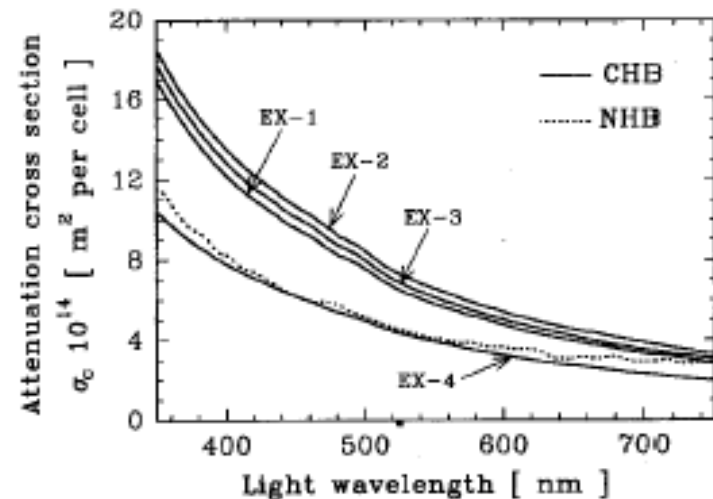
(Stramski and Reynolds 1993)

Optical variability for heterotrophic bacteria

Absorption



Beam attenuation

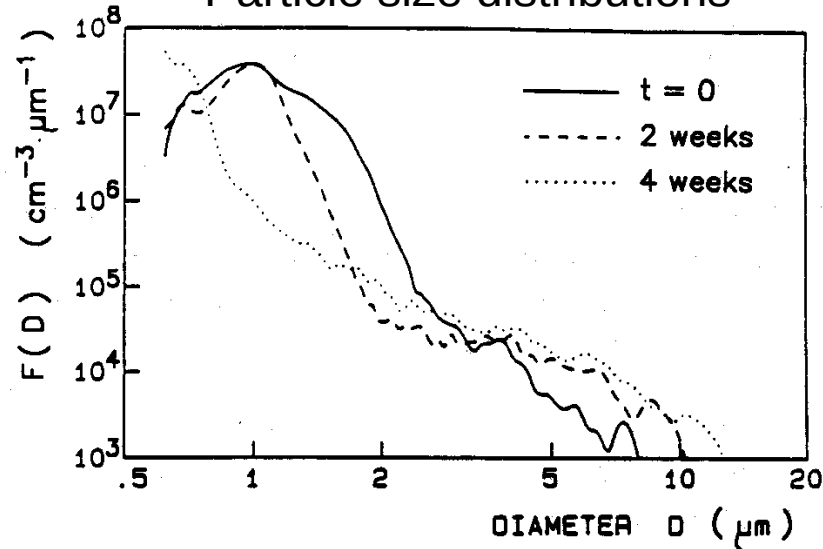


CHB Carotenoid-rich bacteria:
grown in nutrient-enriched seawater [EX-1
(light-dark cycle), EX-2 and EX-3 (dark)],
and in nutrient-poor seawater (EX-4)

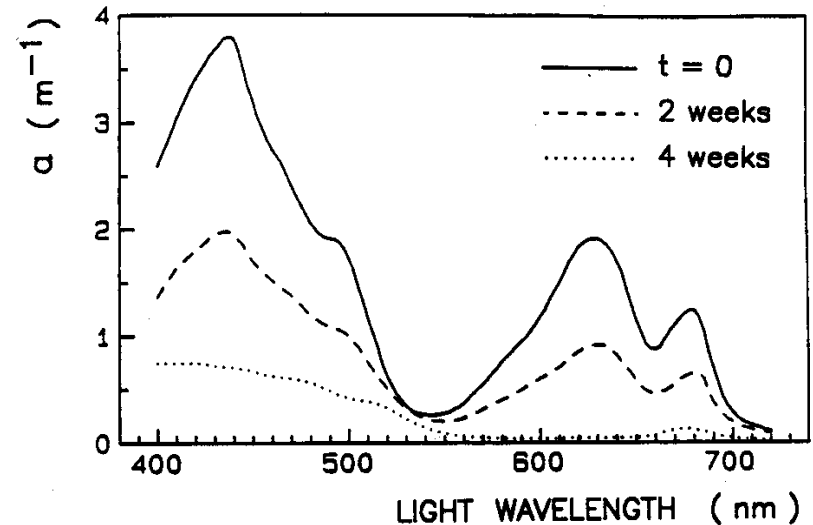
NHB Non-pigmented bacteria:
fast-growing in the absorption experiment
and starved in the attenuation experiment

Prey-predator interactions (cyanobacteria and ciliates)

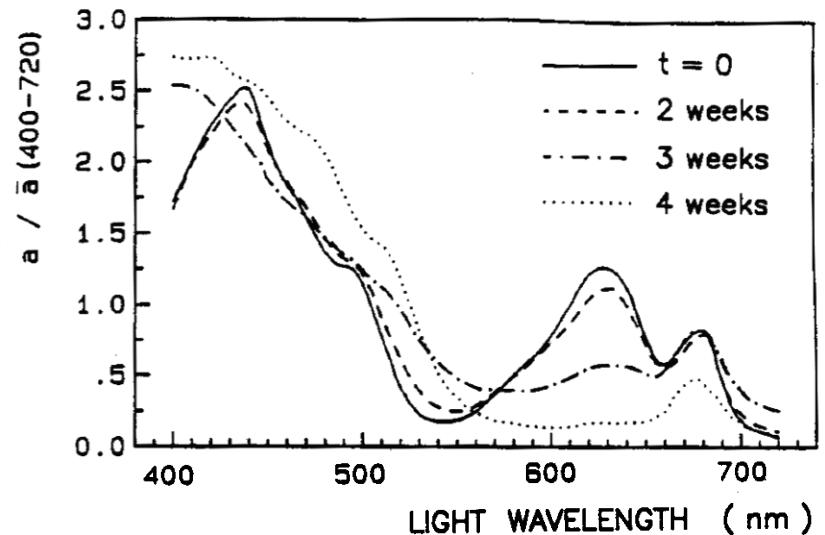
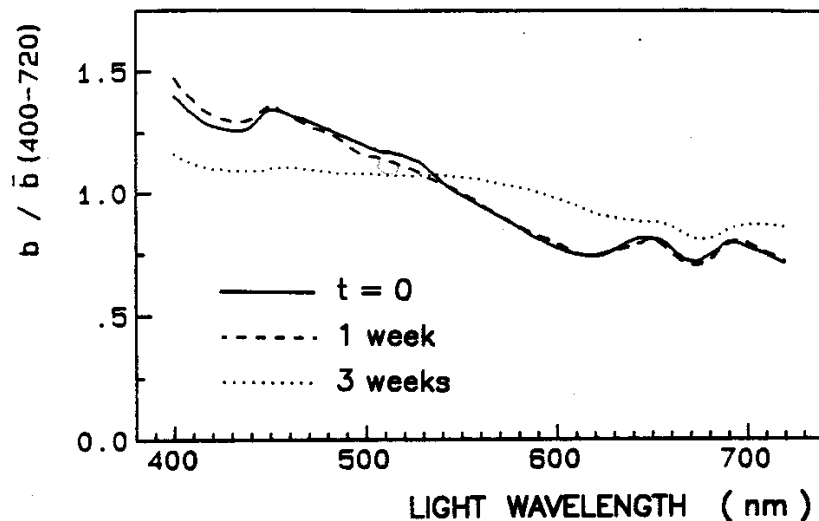
Particle size distributions



Absorption spectra

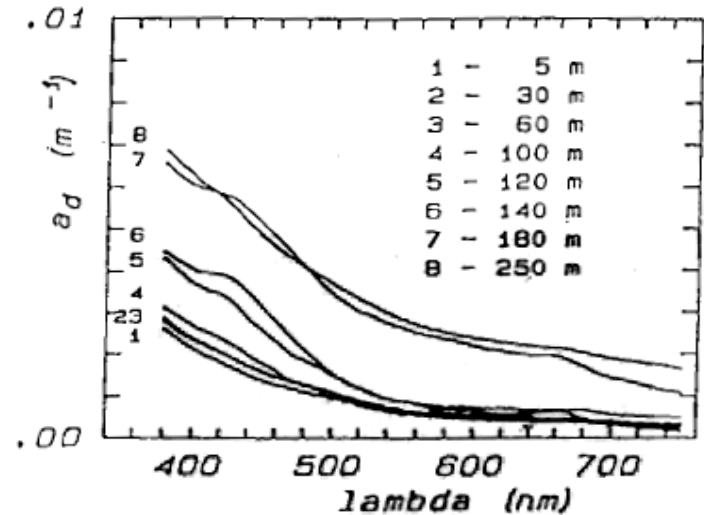
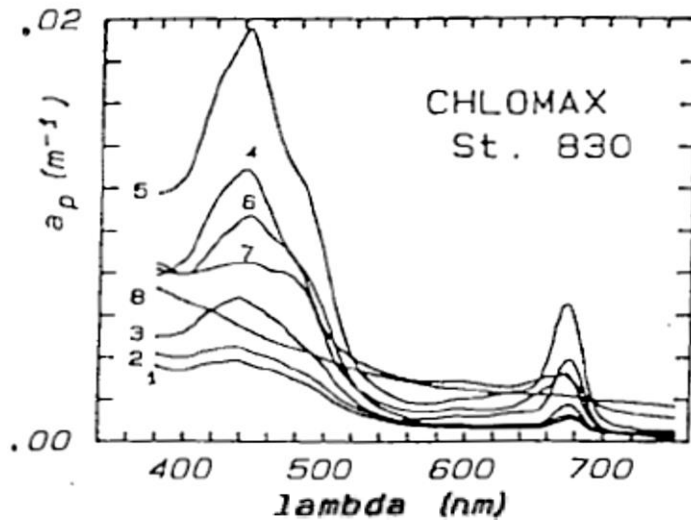


Scattering spectra

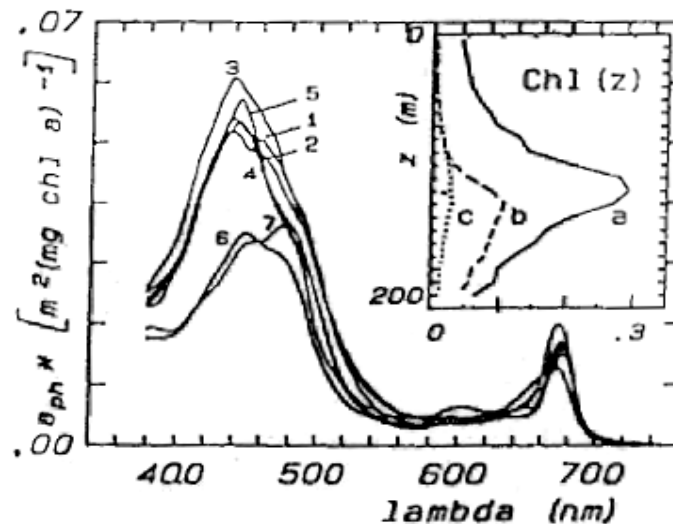


Examples of particulate absorption coefficients

a_p , $a_{d \text{ or } NAP}$, a_{ph} (data from the Sargasso Sea)

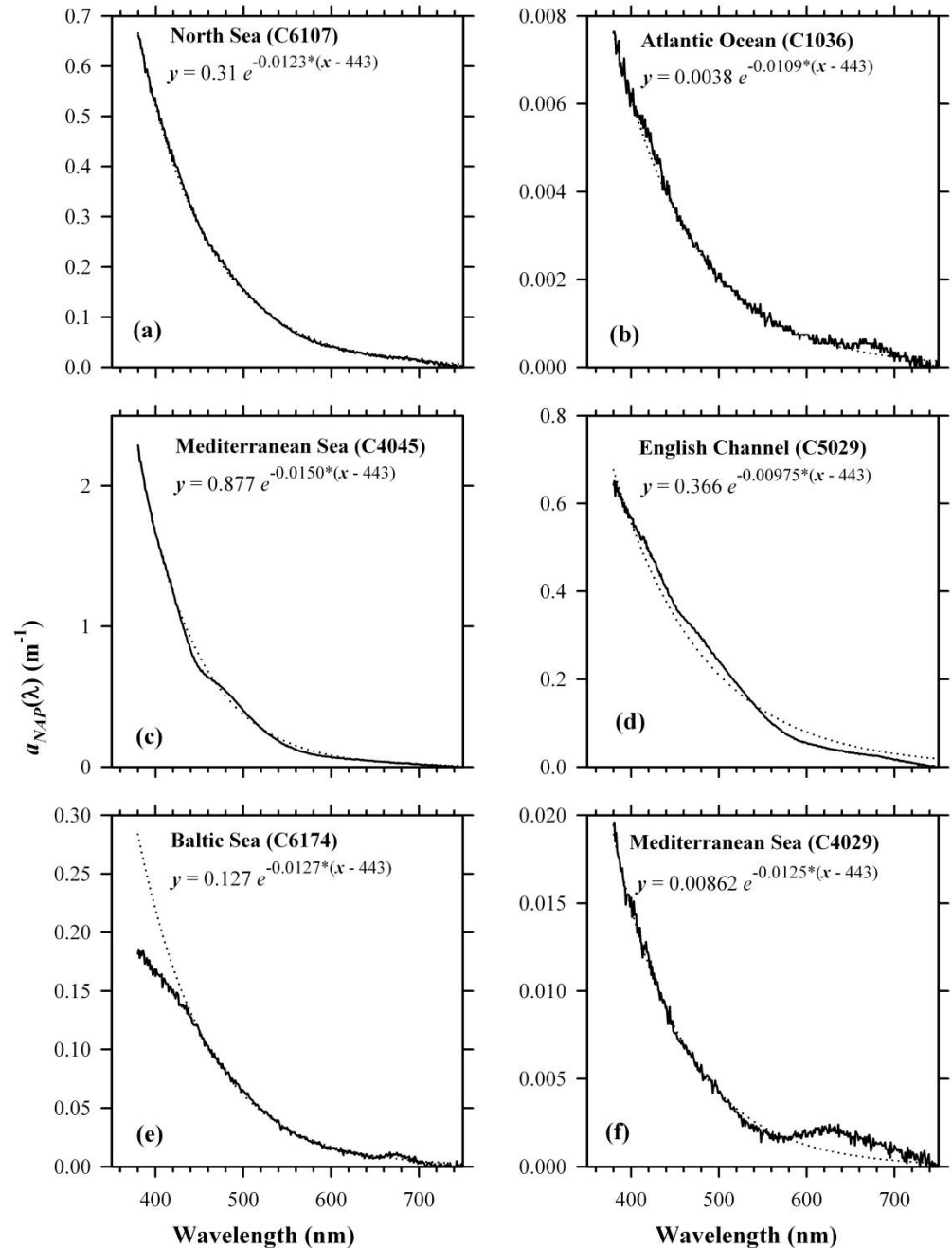


$$a_{ph} = a_p - a_{d \text{ or } NAP}$$



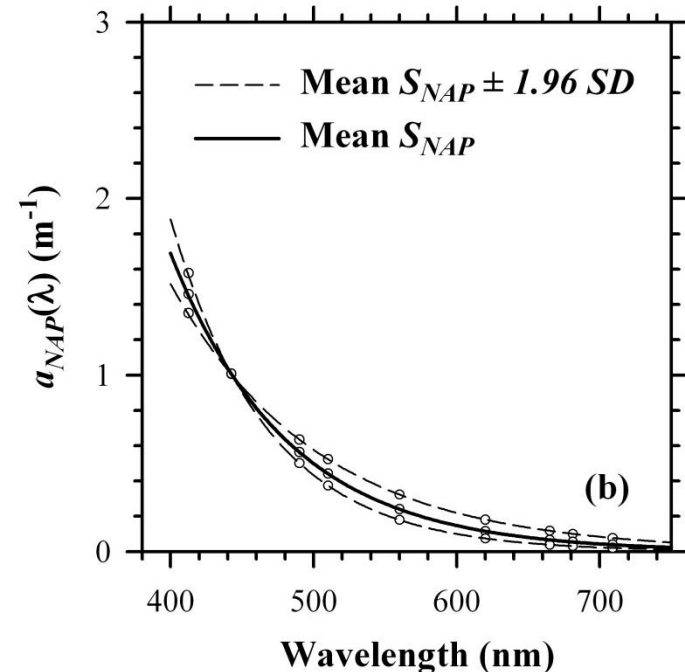
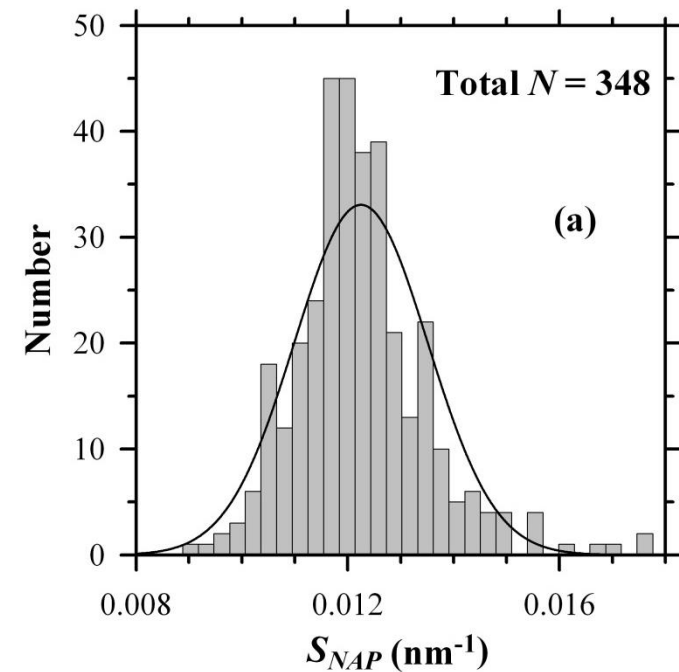
(Bricaud and Stramski 1990)

Example non-algal
particle (NAP)
absorption spectra and
the corresponding
exponential fits for
different regions



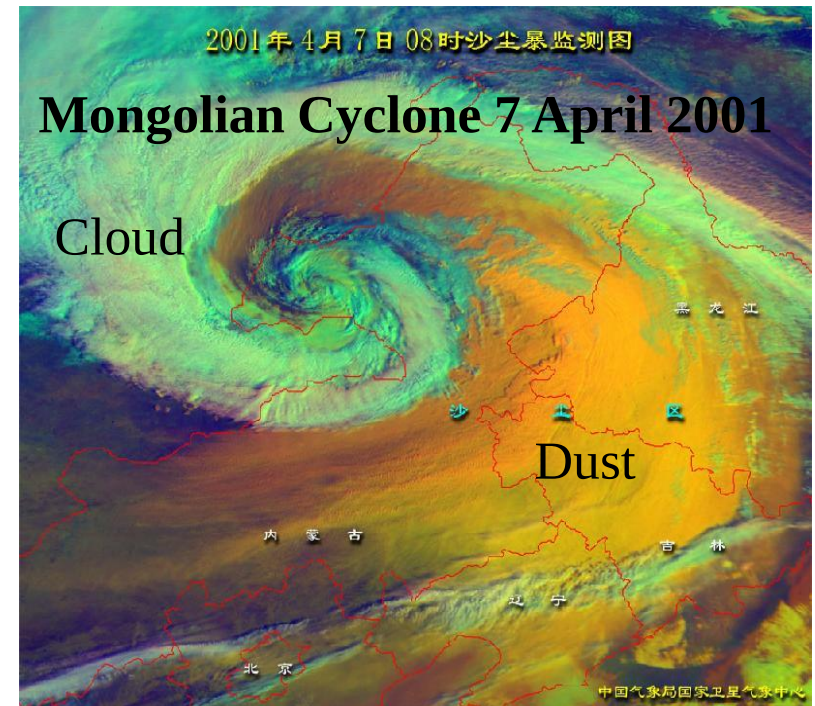
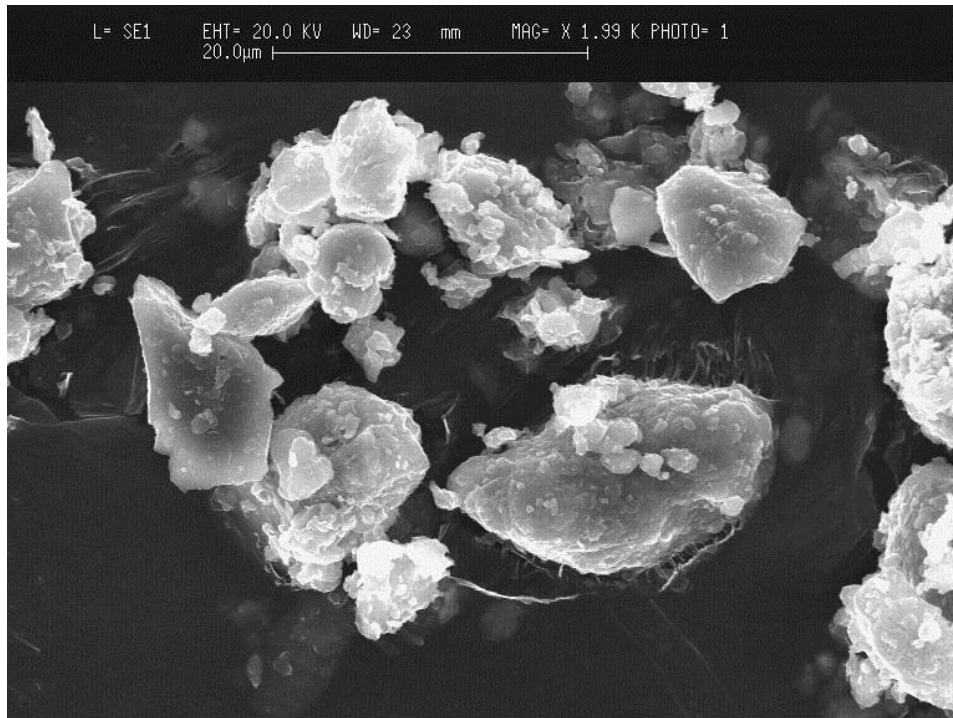
Frequency distribution of spectral slope of NAP absorption

NAP absorption spectra calculated with $a_{NAP}(443)=1 \text{ m}^{-1}$ and $S_{NAP} = 0.0123 \text{ nm}^{-1}$ (± 1.96 standard deviation, where $SD=0.0013 \text{ nm}^{-1}$)



Mineral particles

Saharan dust

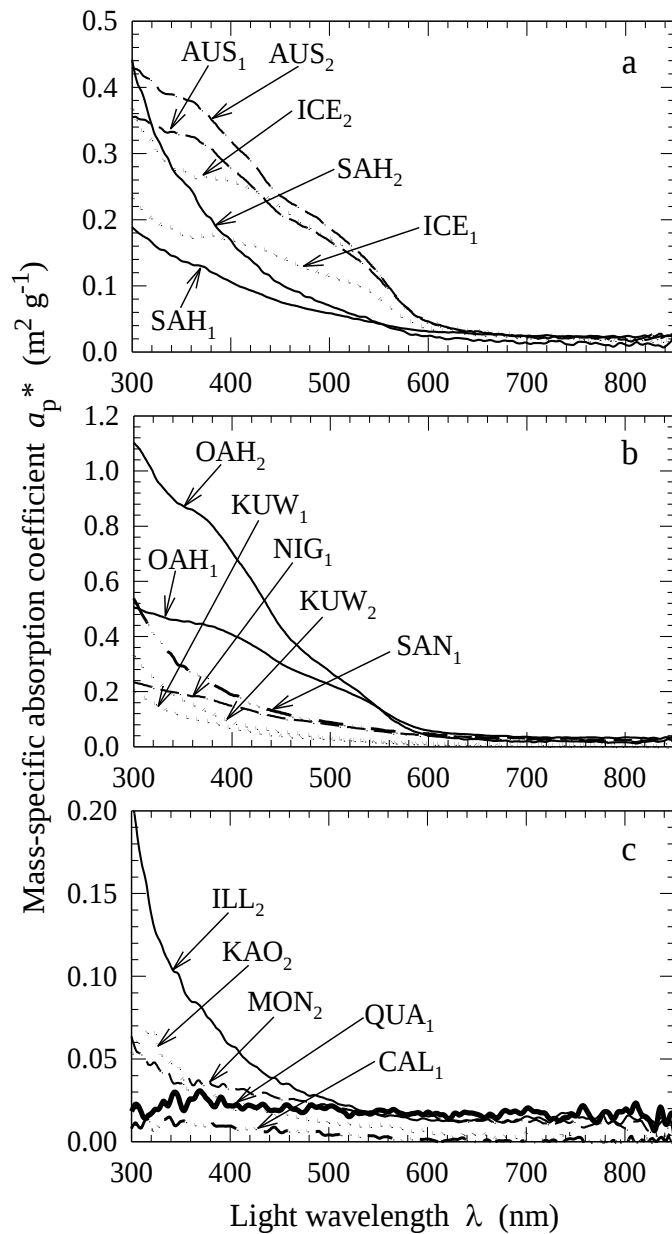


Terrigenous mineral-rich particulate matter

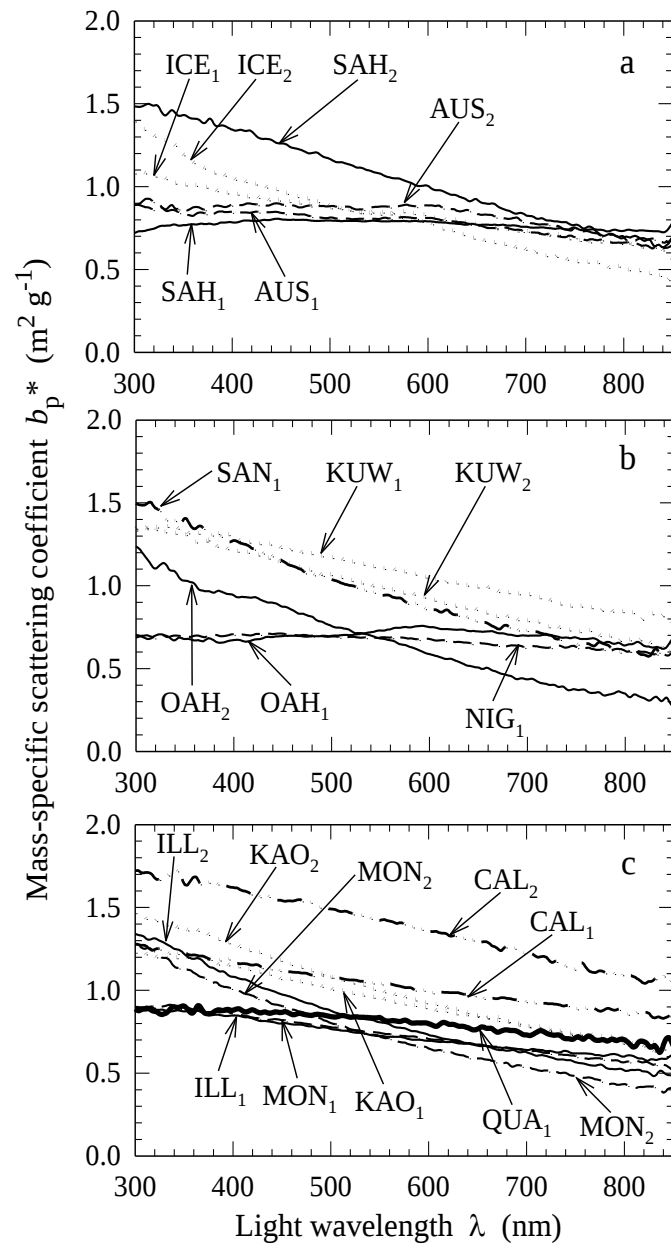
Sample ID	Description	Origin
ILL ₁	illite	Source Clay Minerals Repository, University of Missouri (ref. IMt-1)
ILL ₂	as above but different PSD	as above
KAO ₁	kaolinite (poorly crystallized)	as above (ref. KGa-2)
KAO ₂	as above but different PSD	as above
MON ₁	Ca-montmorillonite	as above (ref. SAz-1)
MON ₂	as above but different PSD	as above
CAL ₁	calcite	natural crystal
CAL ₂	as above but different PSD	as above
QUA ₁	quartz	natural crystal
SAH ₁	atmospheric dust from Sahara	red rain event, Villefranche-sur-Mer, France
SAH ₂	as above but different PSD	as above
AUS ₁	surface soil dust	cliff shore, Palm Beach near Sydney, Australia
AUS ₂	as above but different PSD	as above
ICE ₁	ice-rafted particles	glacier runoff, Kongsfjord, Spitsbergen
ICE ₂	as above but different PSD	as above
OAH ₁	surface soil dust	Oahu, Hawaii Islands
OAH ₂	as above but different PSD	as above
KUW ₁	surface soil dust	Kuwait (eastern part, close to ocean)
KUW ₂	as above but different PSD	as above
NIG ₁	surface soil dust	southwest Nigeria
SAN ₁	atmospheric dust	San Diego, California

(Stramski et al. 2007)

Mass-specific absorption

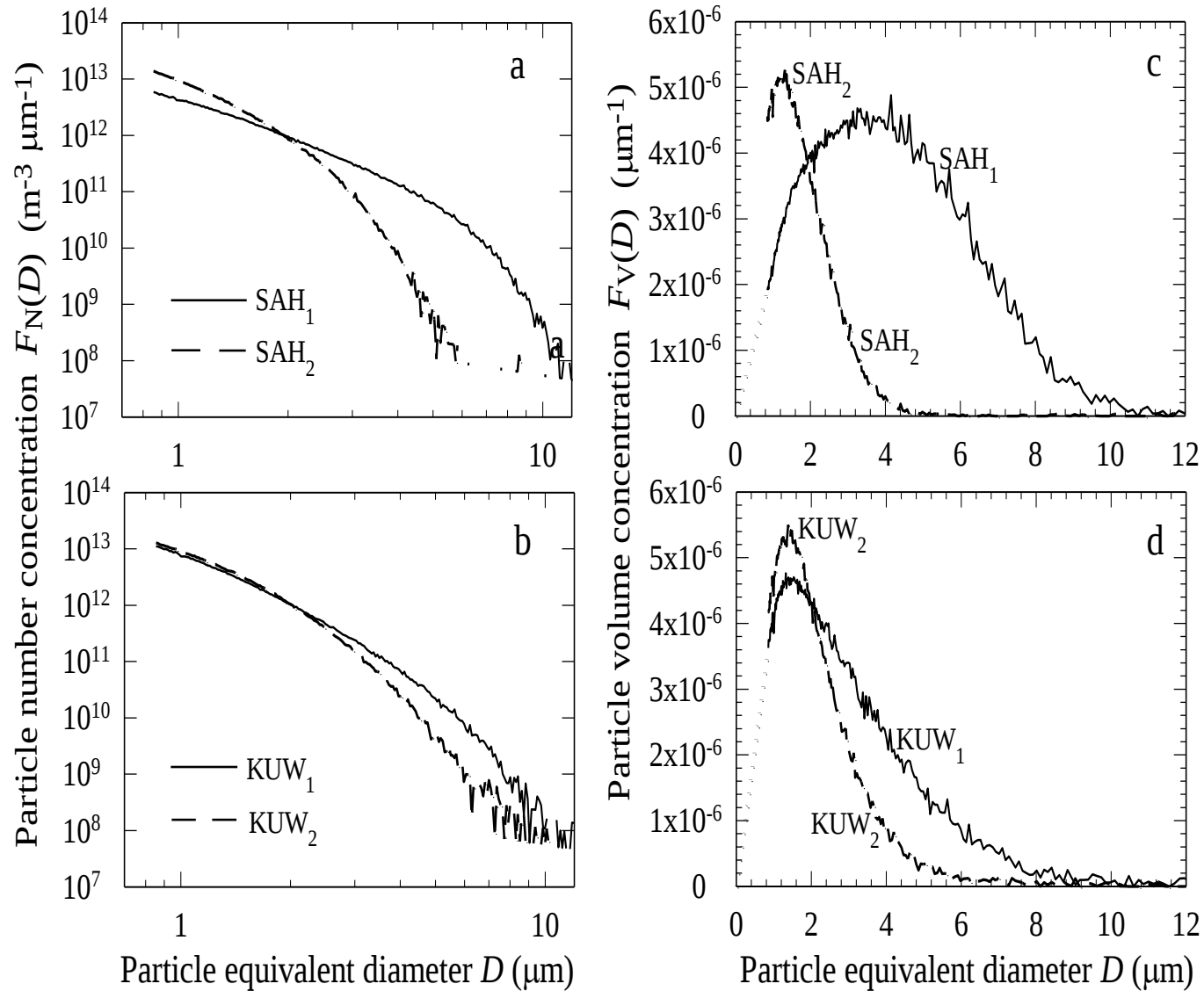


Mass-specific scattering



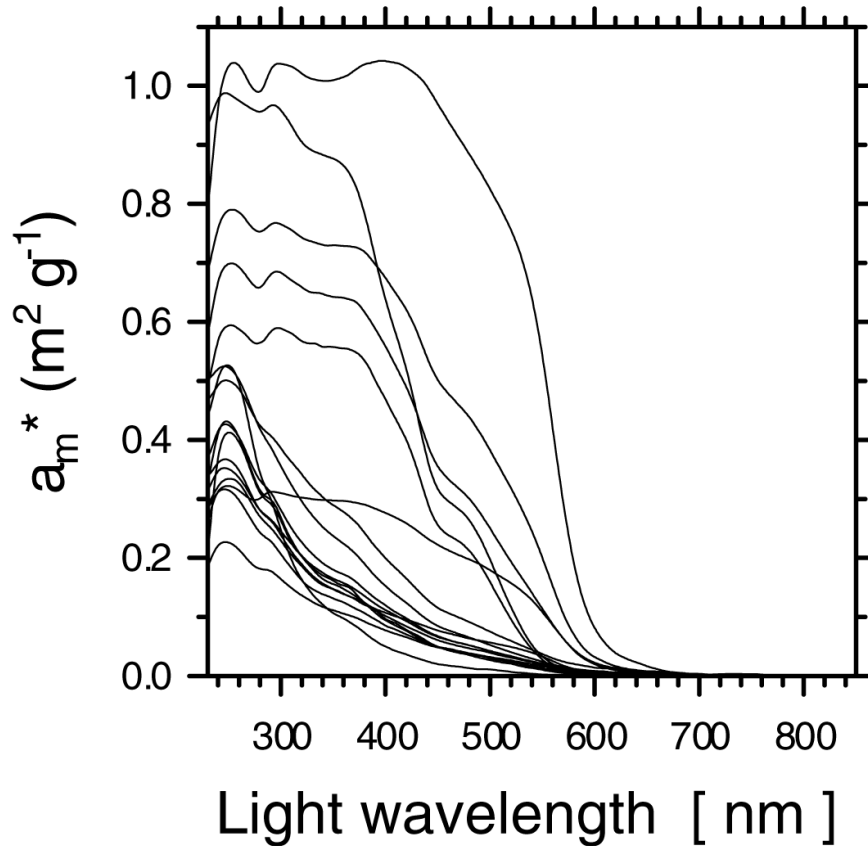
(Stramski et al. 2007)

Particle Size Distributions

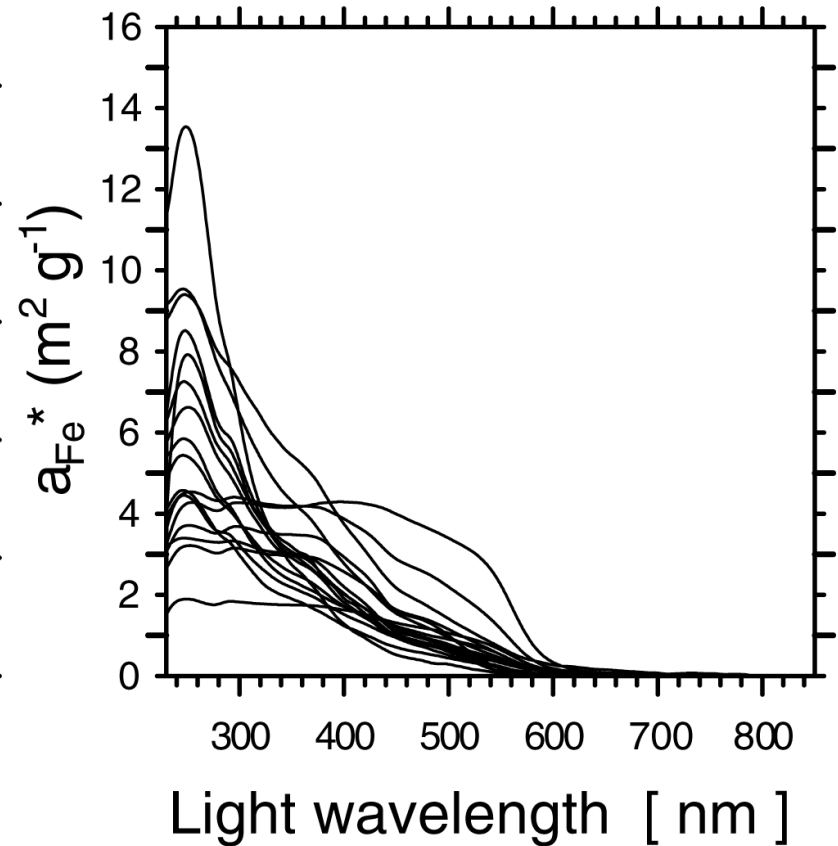


Absorption of mineral-rich particulate assemblages

Mass-specific absorption



Fe-specific absorption





ICE



R36



M1



ALG1



Fe(OH)₃



AUS



R37



M2



ALG2



SAH1



ILL2



R38



M3



NIG1



SAH2



CAL



R39



M4

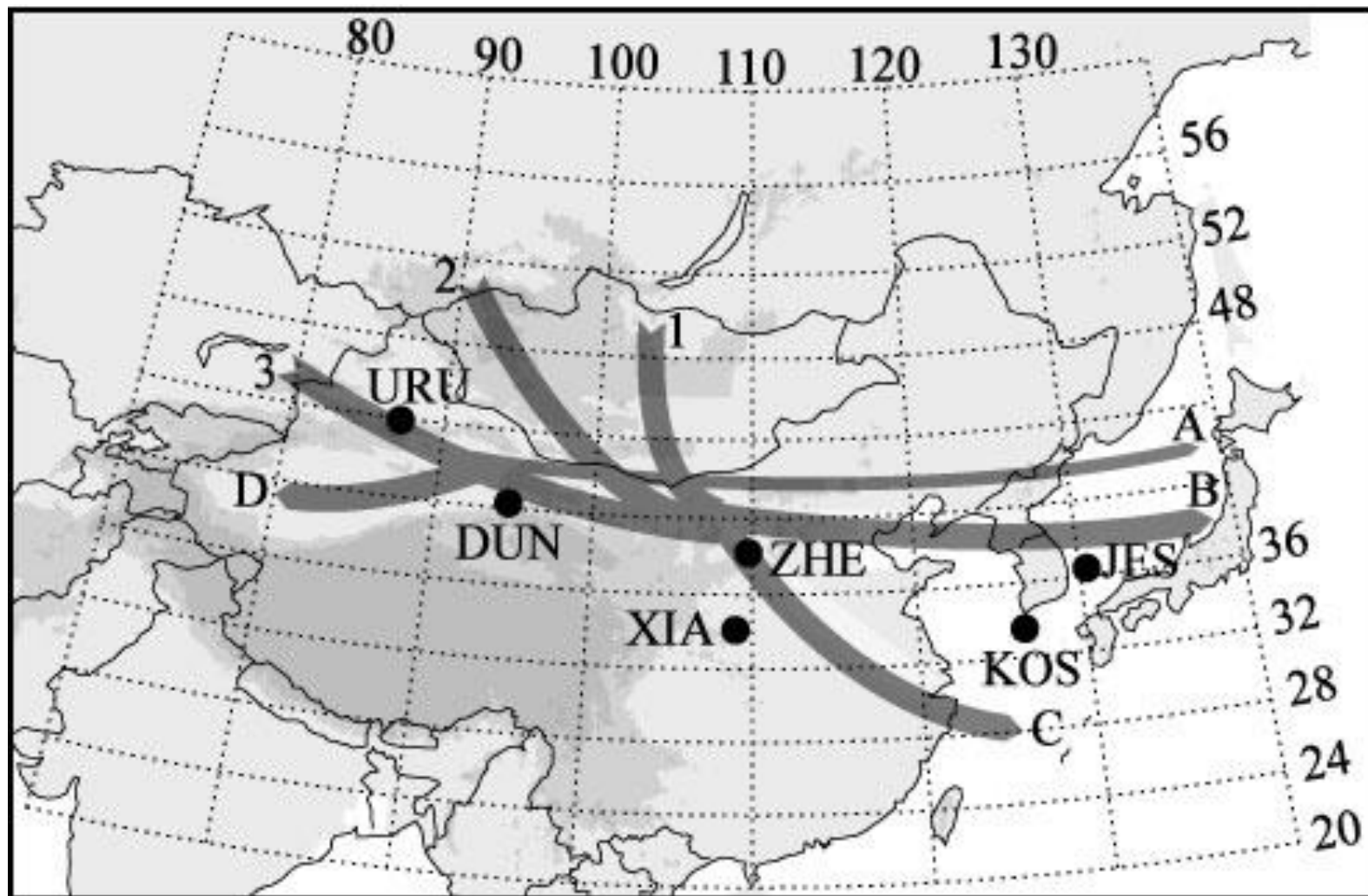


NIG2

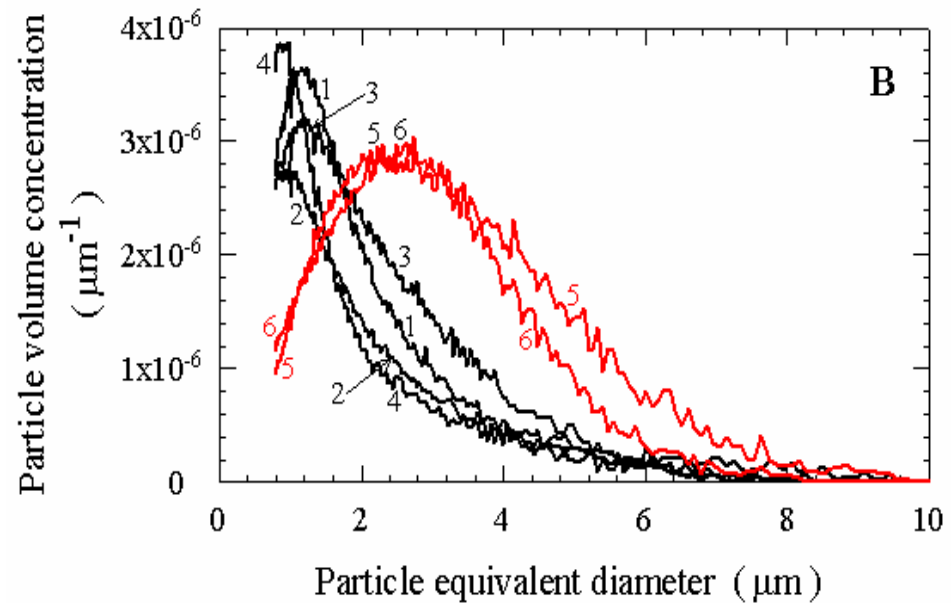
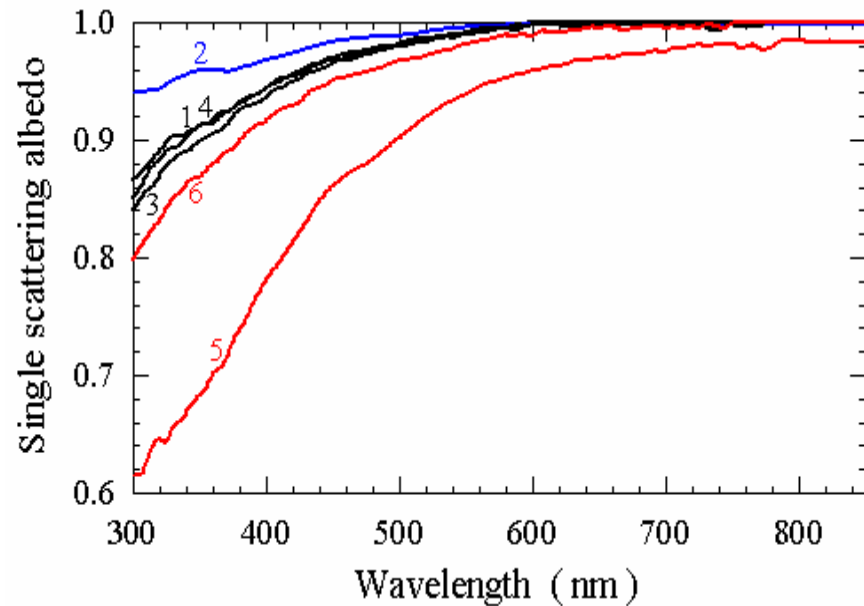
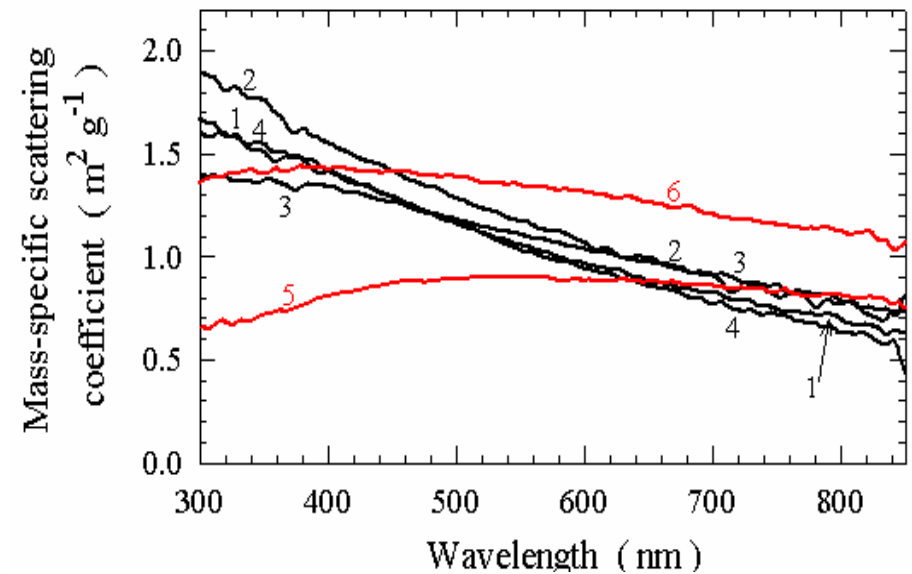
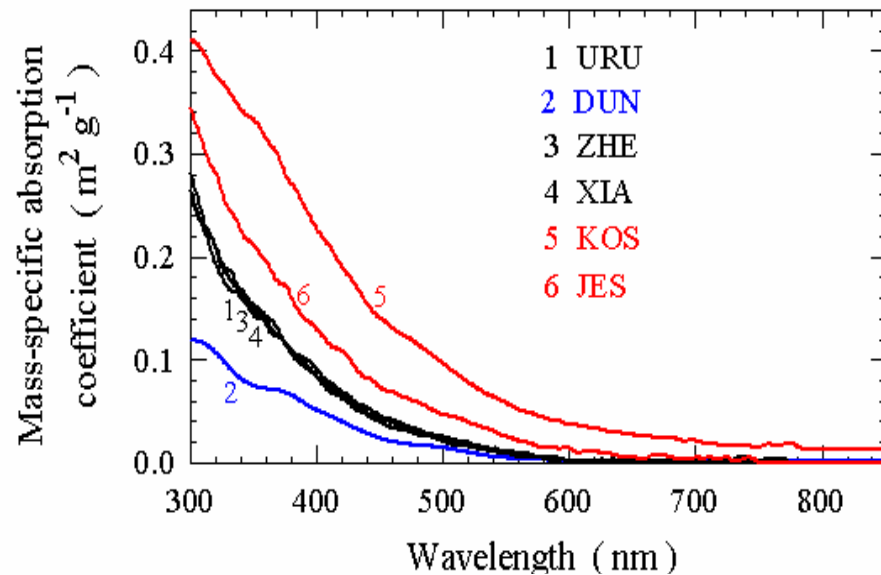


KUW

Asian dust - sampling locations



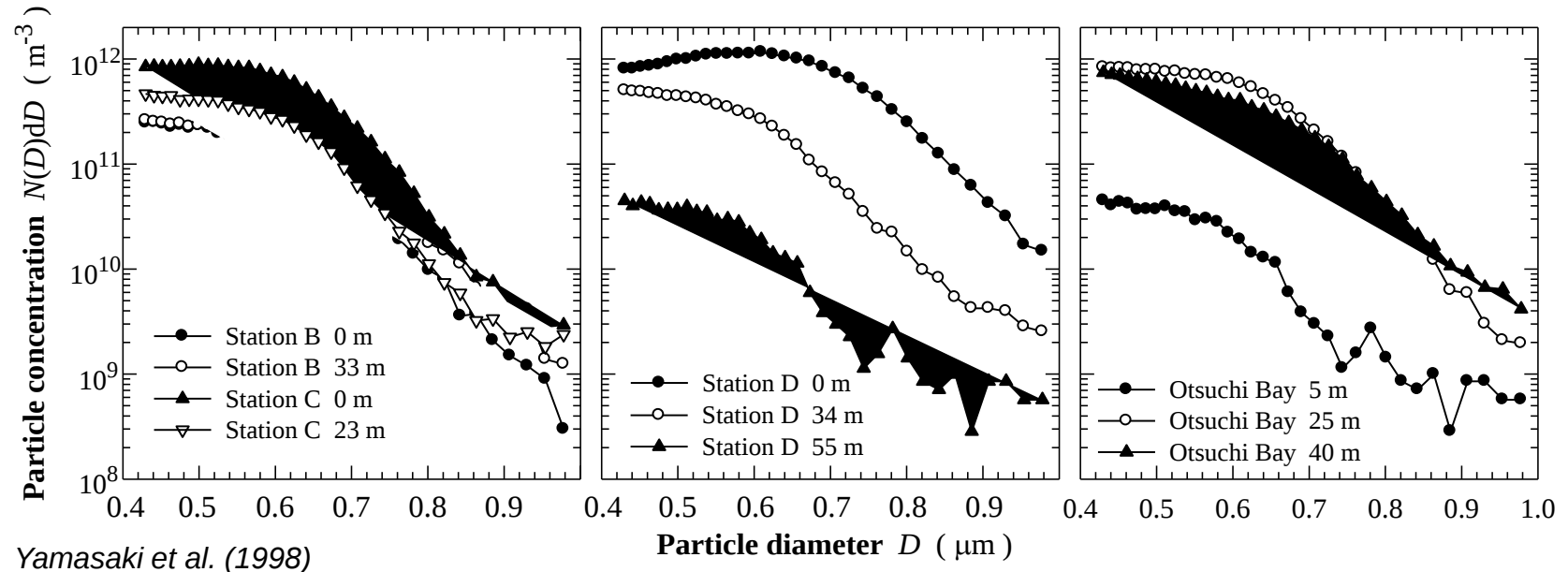
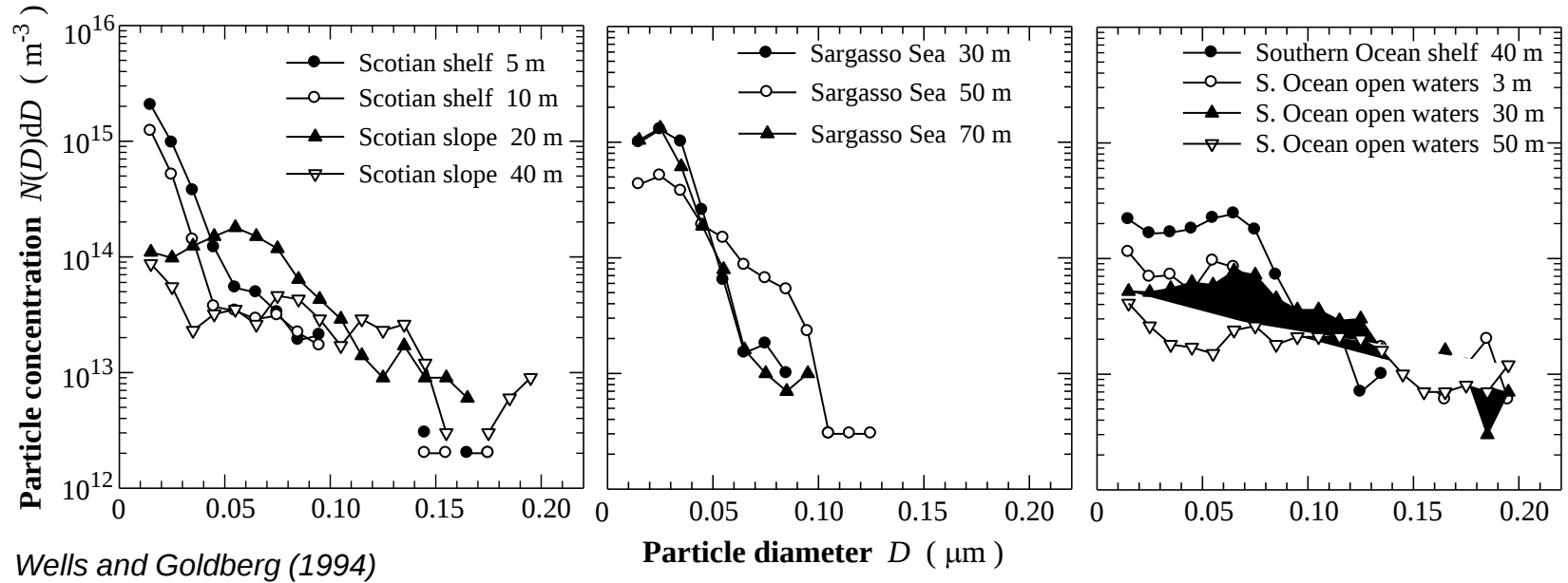
Asian mineral-rich dust



Colloidal particles ($< 1\ \mu\text{m}$ in size) undergoing Brownian motion

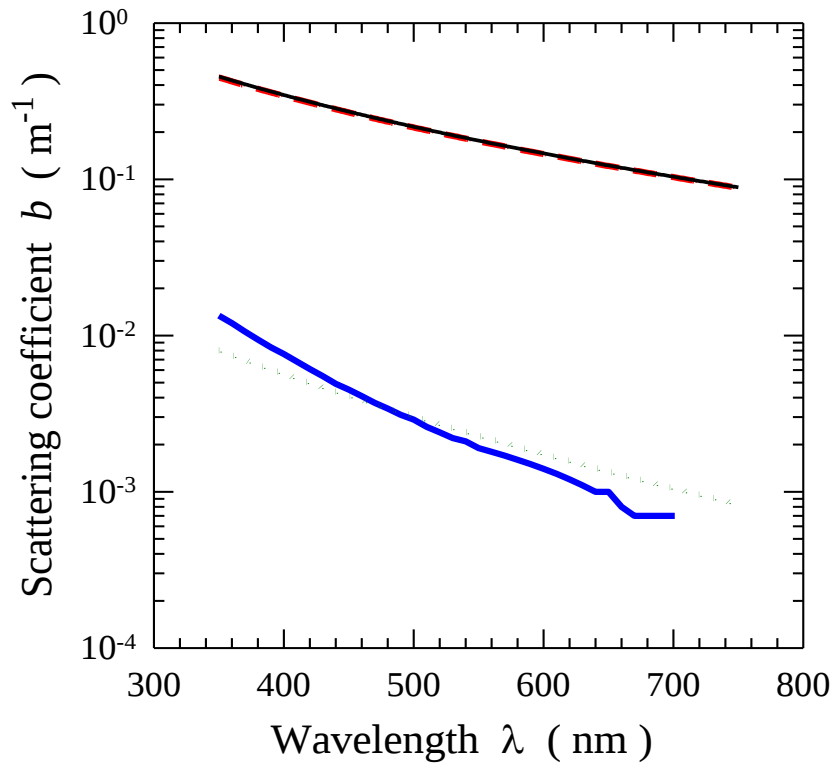


Size distributions of colloids

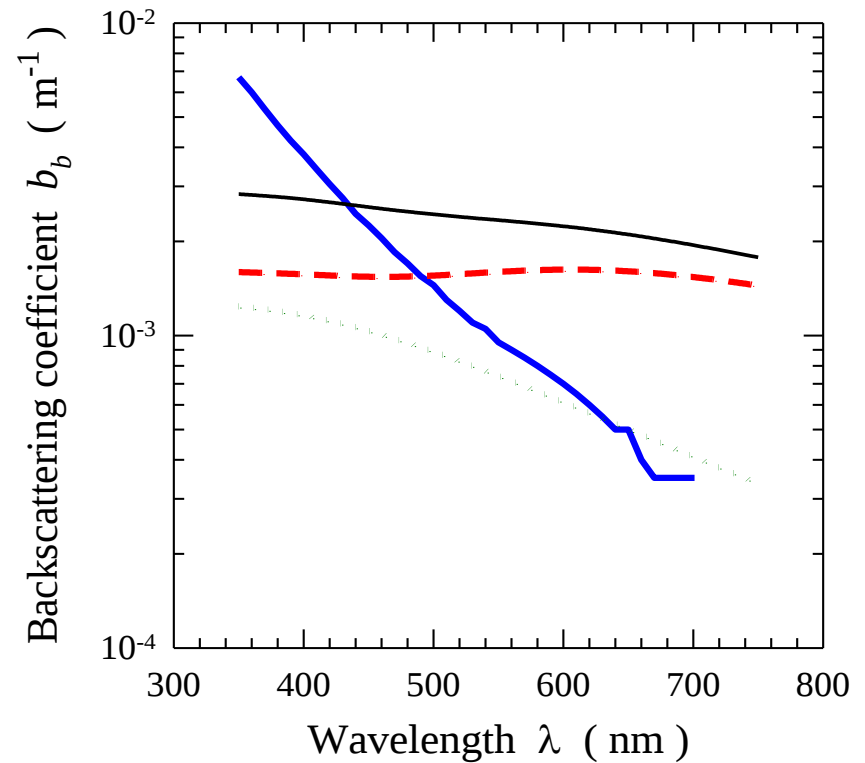


Results for colloidal particles

scattering

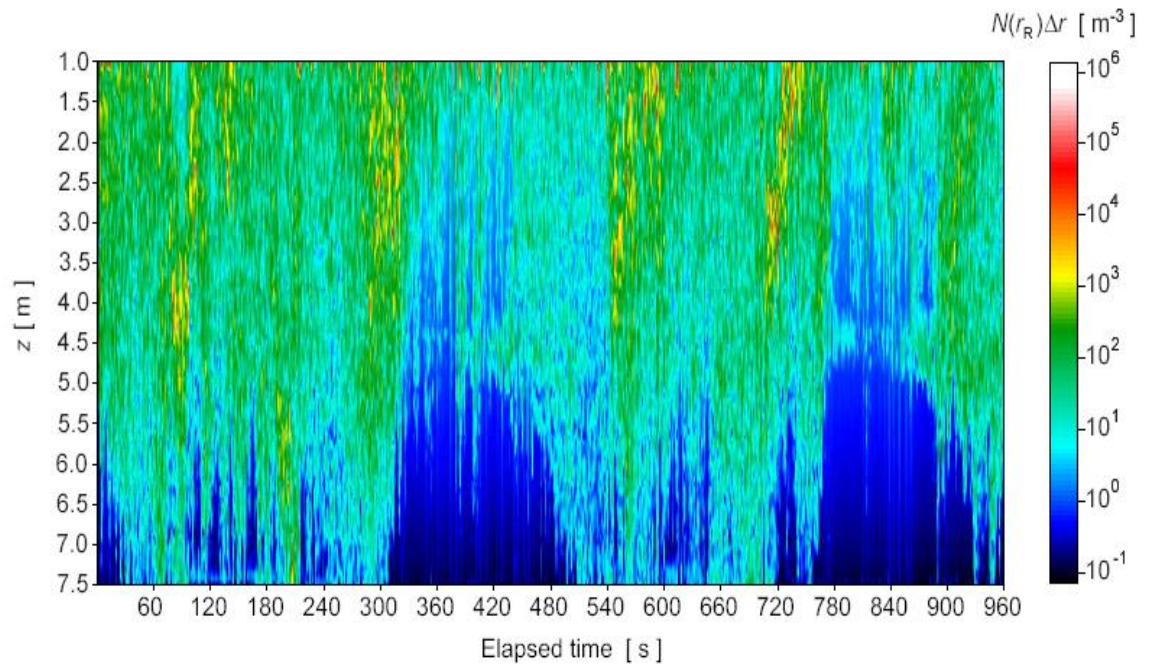


backscattering



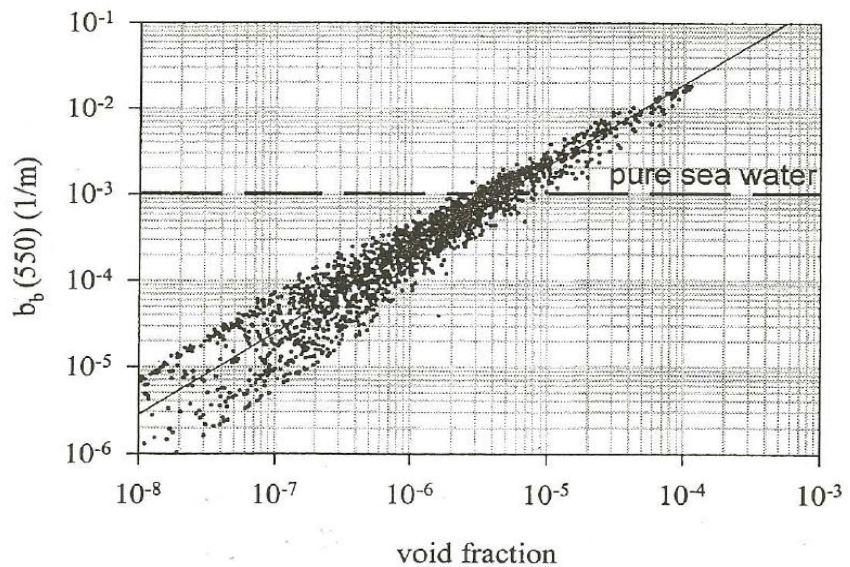
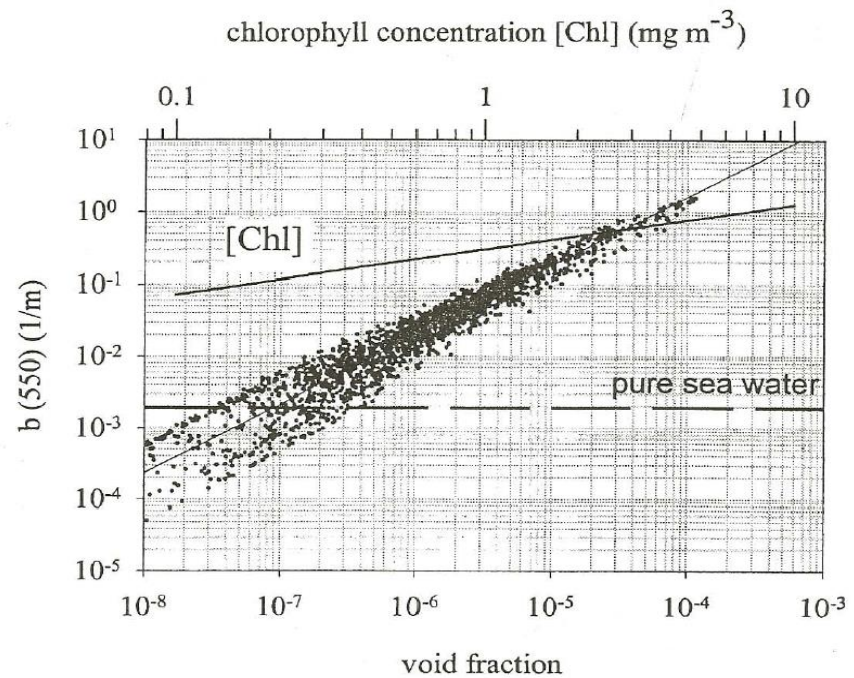
- pure seawater
- ... average small colloids
- - - average large colloids
- small + large colloids

Light scattering by bubbles entrained by wave breaking



(Stramski and Tęgowski 2001)

Scattering and backscattering by bubbles as a function of void fraction



(Terrill et al. 2001)

Traditional approach

Inherent Optical Properties (IOPs) described in terms of a few broadly-defined categories of seawater constituents

$$IOP(\lambda) = IOP_w(\lambda) + IOP_p(\lambda) + IOP_{CDOM}(\lambda)$$

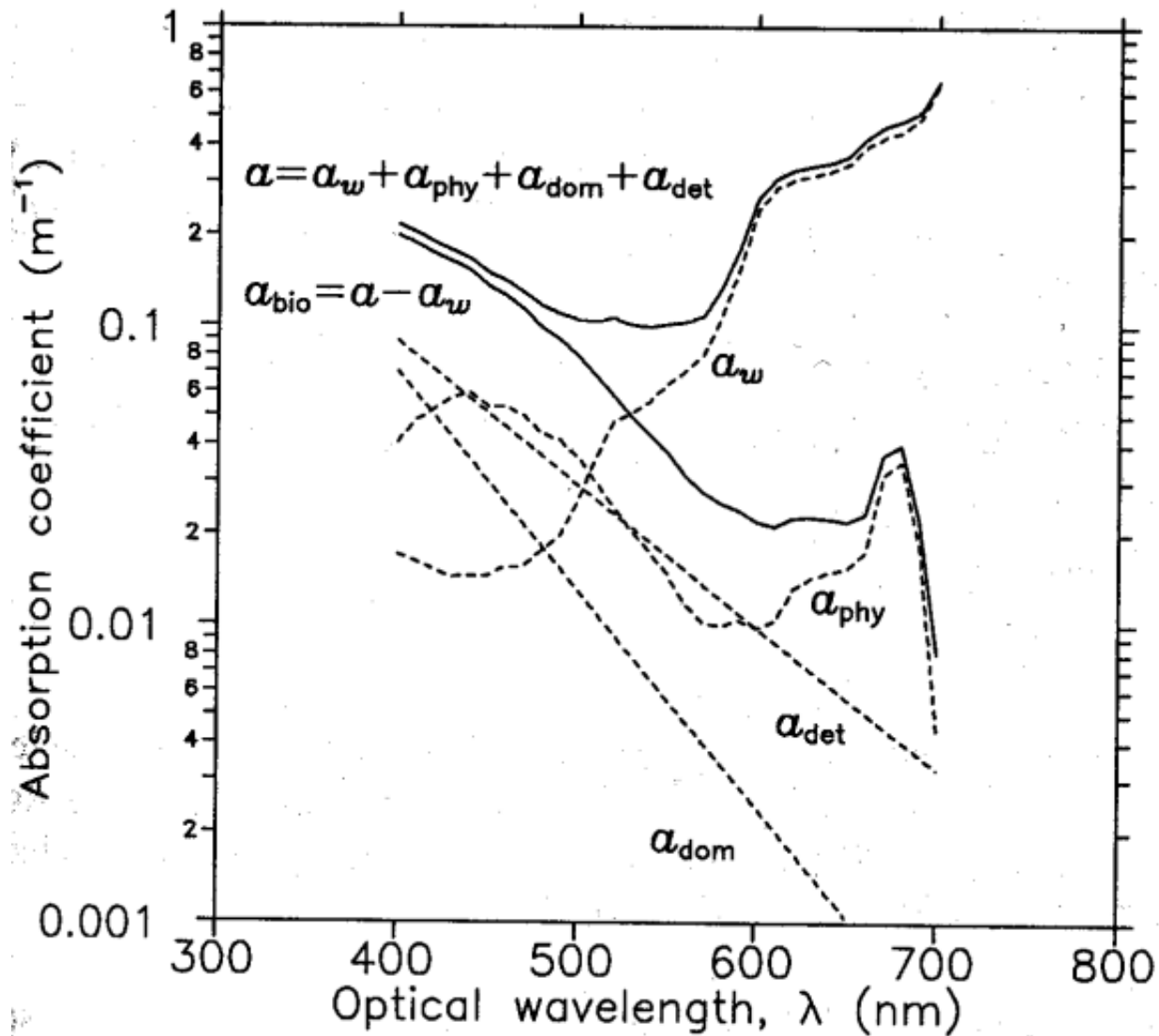
$$IOP_p(\lambda) = IOP_{ph}(\lambda) + IOP_{NAP}(\lambda)$$

Example IOPs:

absorption coefficient, scattering coefficient,

beam attenuation coefficient, volume scattering function

A four-component model of absorption



Chlorophyll-based approach

$$IOP(\lambda) = IOP_w(\lambda) + f [Chla]$$

$$\text{for example } a_{ph}(\lambda) = f [Chla]$$

$$a_p(\lambda) = f [Chla]$$

$$AOP(\lambda) \text{ (e.g., ocean reflectance)} = f [Chla]$$

Case 1 and Case 2 Waters

CASE 1 WATERS

LIVING ALGAL CELLS
variable concentration

ASSOCIATED DEBRIS
*Originating from grazing by
zooplankton and natural decay*

DISSOLVED ORGANIC MATTER
*liberated by algae and their debris
(yellow substance)*

RESUSPENDED SEDIMENTS
*from bottom along the coastline and
in shallow areas*

TERRIGENOUS PARTICLES
river and glacial runoff

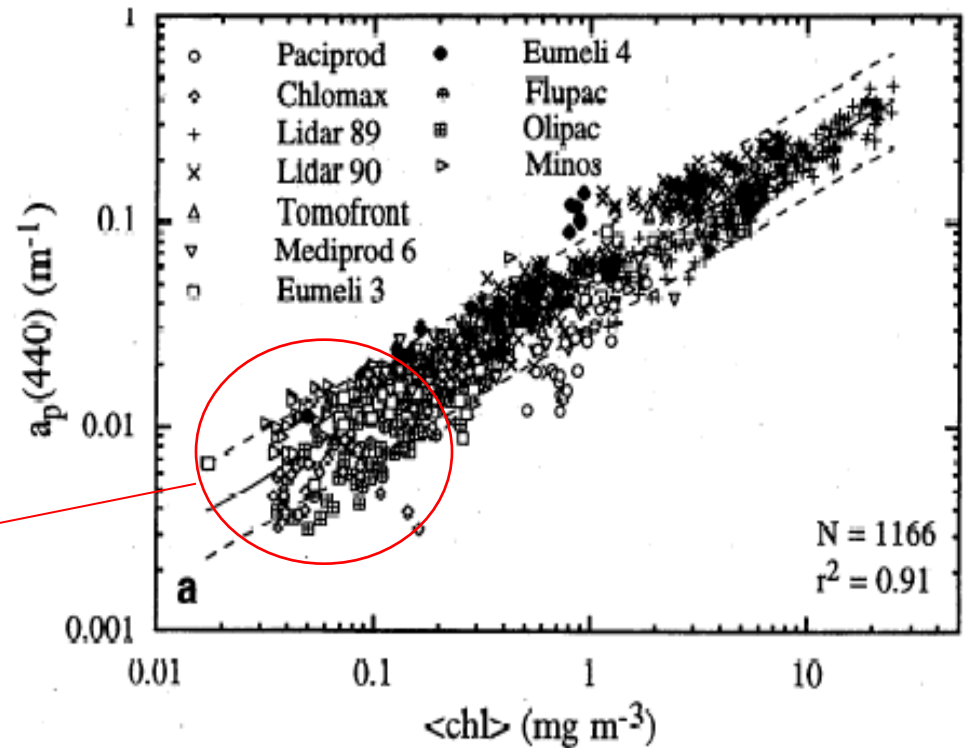
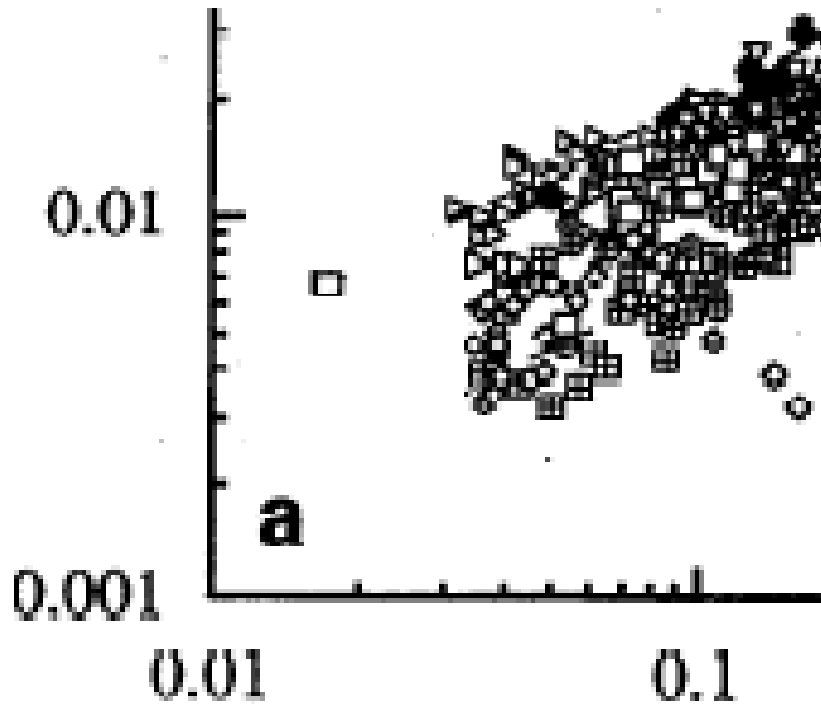
DISSOLVED ORGANIC MATTER
*land drainage (terrigenous yellow
substance)*

ANTHROPOGENIC INFLUX
particulate and dissolved materials

CASE 2 WATERS

Absorption vs. chlorophyll-a

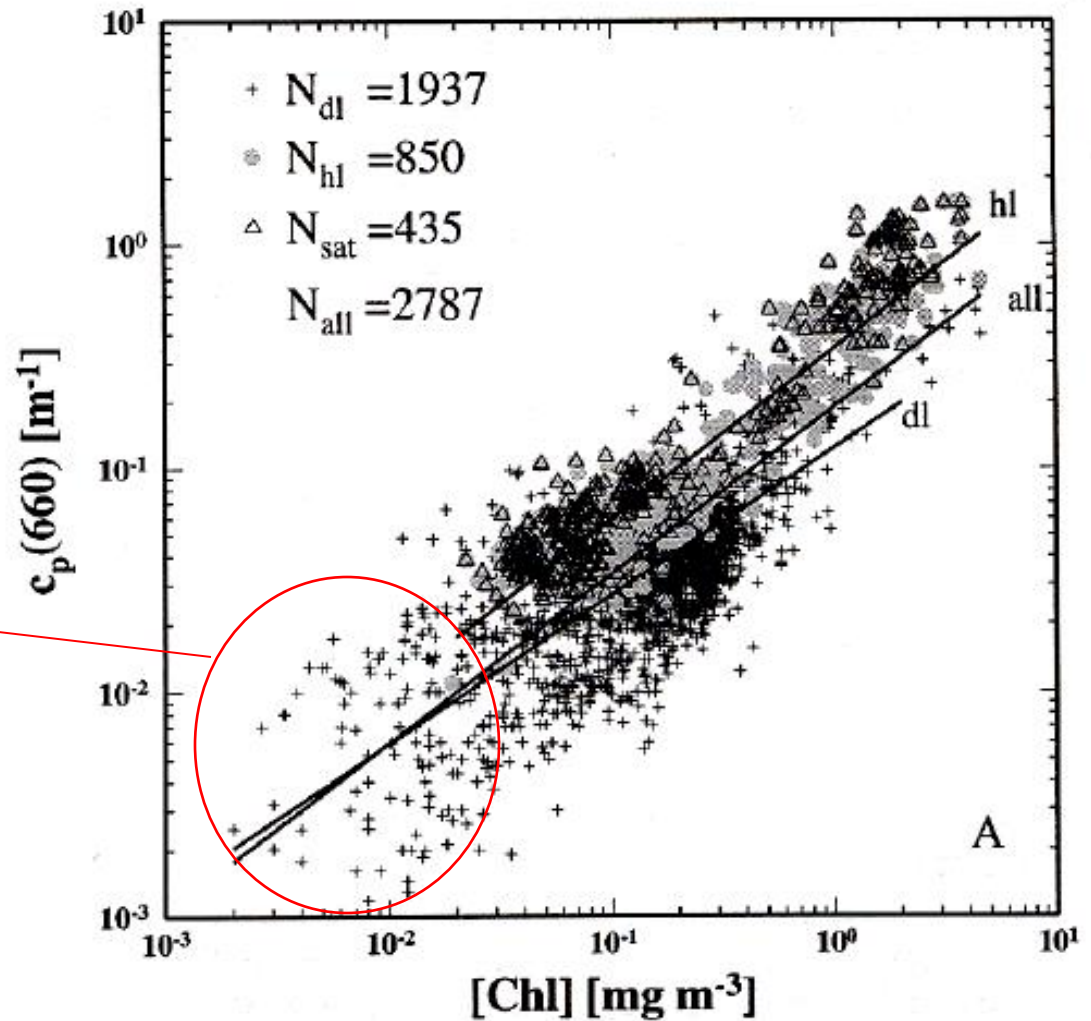
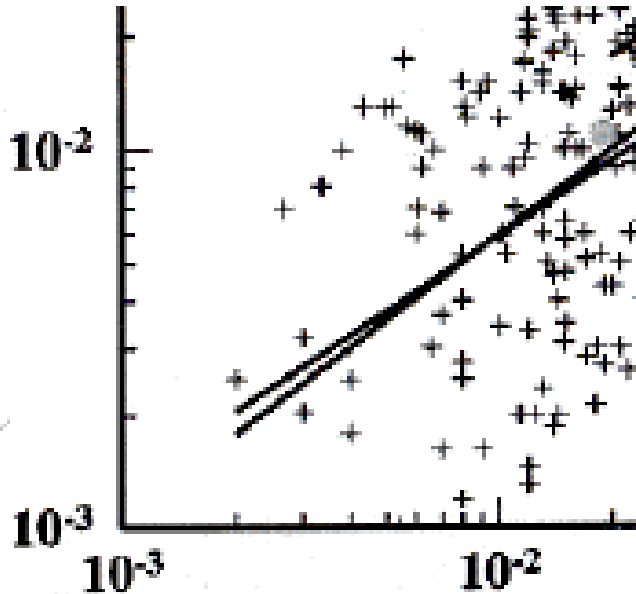
~ 4-fold variation



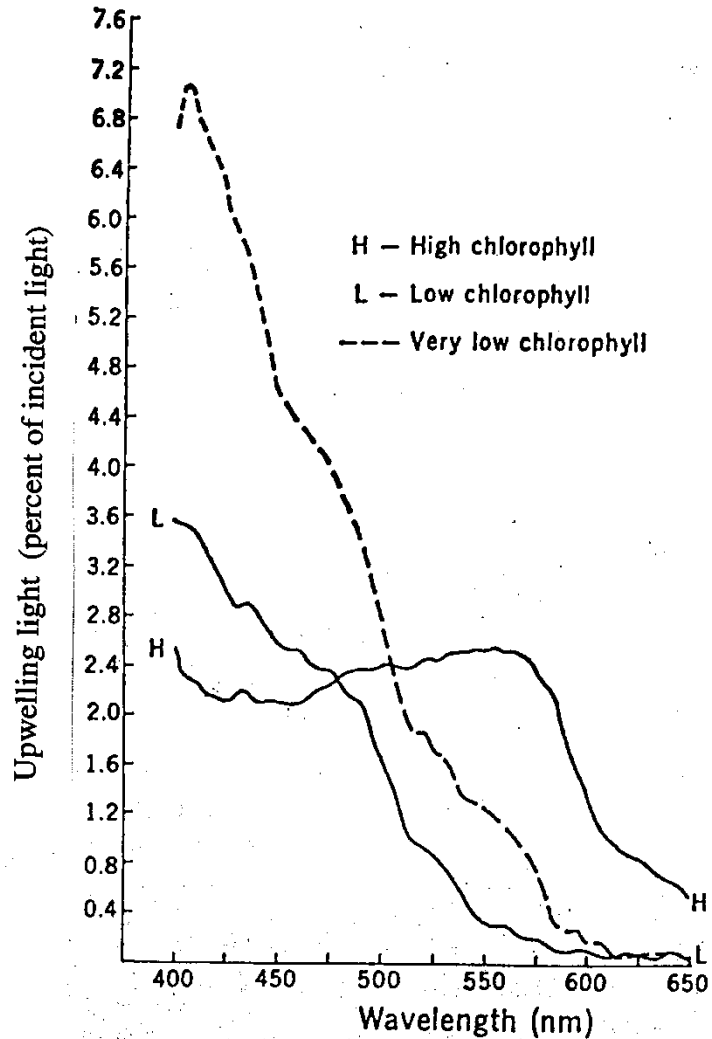
Cruise	Location
PACIPROD	Peru upwelling
CHLOMAX	Sargasso Sea
LIDAR 89	St. Lawrence estuary and gulf
LIDAR 90	St. Lawrence estuary and gulf
TOMOFRONT	northwestern Mediterranean
MEDIPROD 6	southwestern Mediterranean
EUMELI 3	tropical North Atlantic
EUMELI 4	tropical North Atlantic
FLUPAC	equatorial and subequatorial Pacific
OLIPAC	equatorial and subequatorial Pacific
MINOS	eastern and western Mediterranean

Beam attenuation vs. chlorophyll

> 10-fold variation

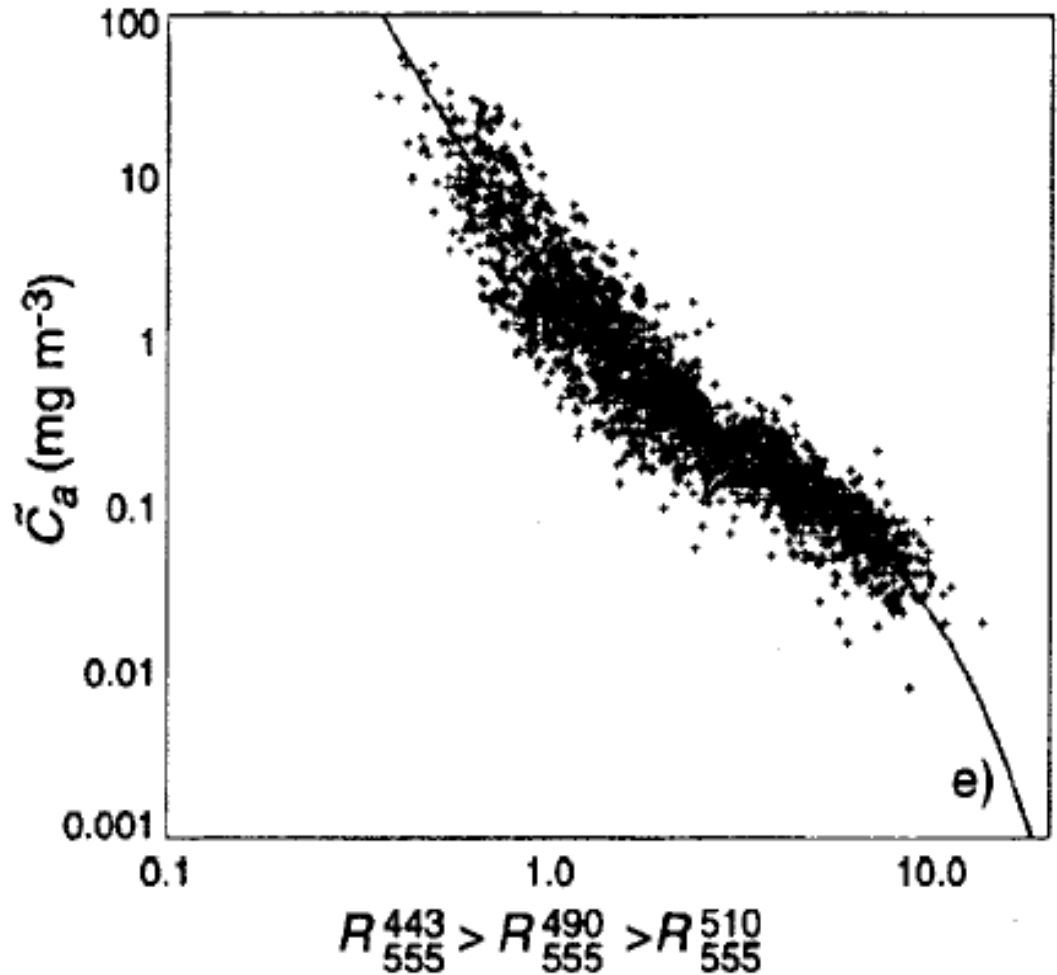


Chlorophyll Algorithm



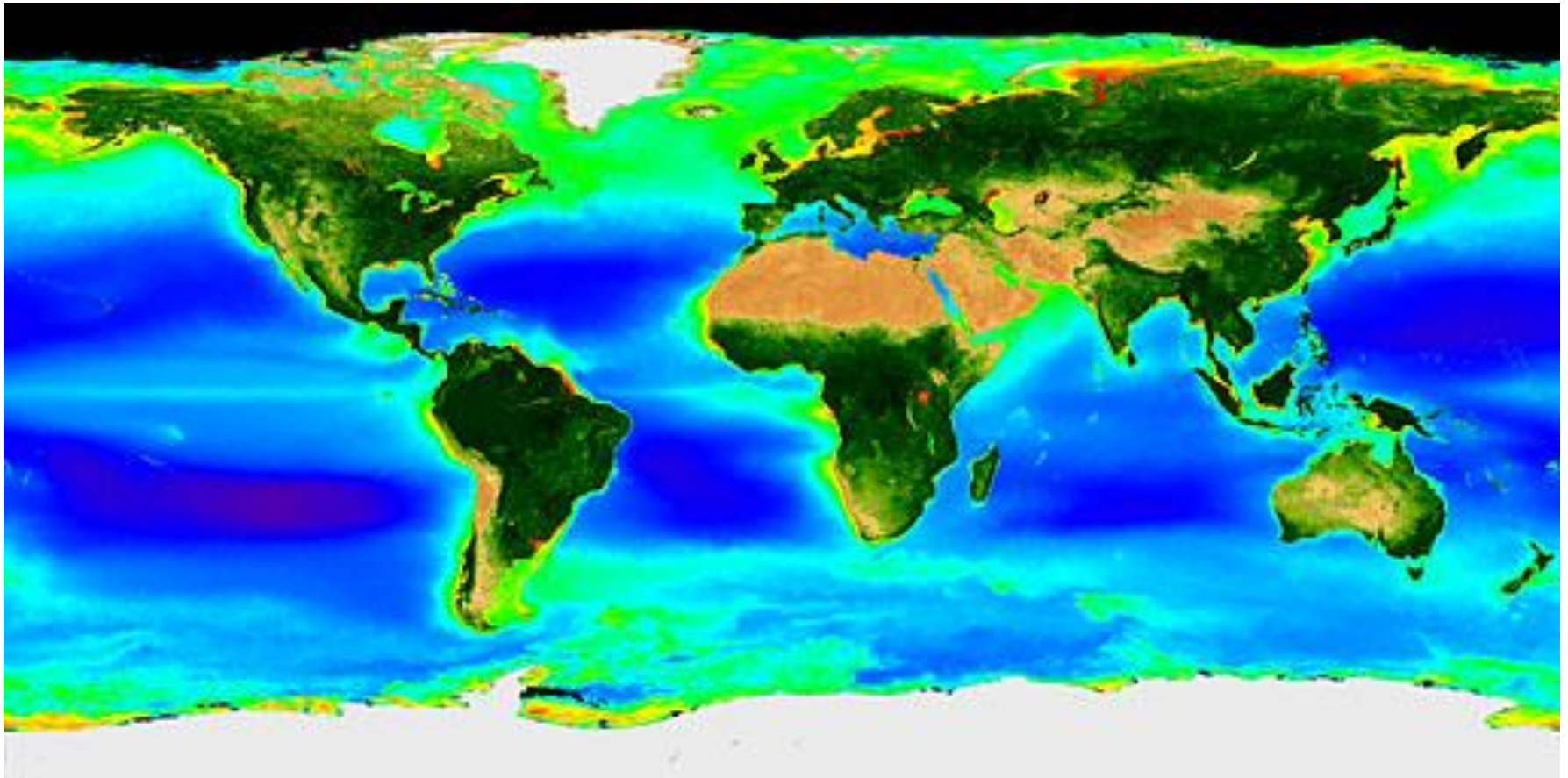
(Clarke, Ewing & Lorenzen 1970)

OC4 Algorithm



(O'Reilly et al. 2000)

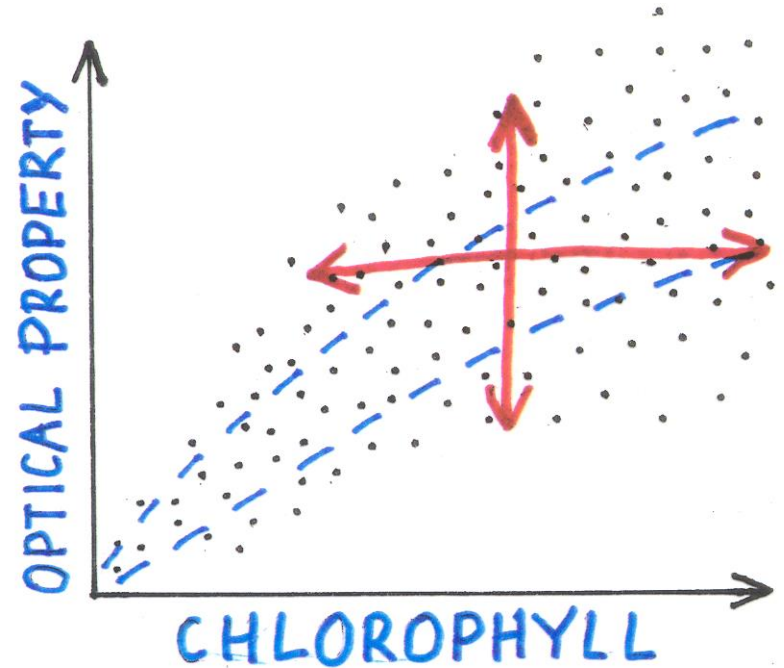
Global distribution of phytoplankton chlorophyll in the world's oceans from Sea-viewing Wide Field-of-view Sensor (SeaWiFS) based on Sep 1997 - Feb 2007 data



Courtesy of NASA

Chlorophyll-based approach: Summary

- Parameterization in terms of chlorophyll-a concentration alone
- Empirical regressions (statistically-derived models)
- Provide average trends but no information about variability
- Not valid for Case 2 waters
- Not necessarily satisfactory for Case 1 waters



Reductionist approach

To develop an understanding and assemble a model of the whole, from the reductionist study of its parts

$$\begin{aligned} IOP_p(\lambda) &= \sum_k IOP_{k,pla}(\lambda) && \text{plankton} \\ &+ \sum_m IOP_{m,min}(\lambda) && \text{minerals} \\ &+ \sum_n IOP_{n,det}(\lambda) && \text{detritus} \end{aligned}$$

Example criteria

- Manageable number of components
- The sum of components should account for the total bulk particulate and optical properties as accurately as possible
- The components should play specific well-defined roles in ocean optics and biogeochemistry

Example reductionist model of particle functional types (PFTs)

Living Particles

Autotrophs

1. Picophytoplankton <2-3 μm
(prokaryotes and eukaryotes)
2. Small nanophytoplankton ~2-8 μm
3. Coccolithophores
4. Large nanophytoplankton ~8-20 μm
5. Microphytoplankton 20-200 μm

Heterotrophs

6. Bacteria ~0.5 μm
7. Microzooplankton O(1-100) μm

Non-Living Particles

Organic

8. Small colloids 0.02-0.2 μm
9. Coarse colloids 0.2-1 μm
10. Detritus >1 μm

Inorganic

11. Colloidal/Clay minerals
<2 μm
12. Larger (Silt and Sand-sized) minerals >2 μm

Challenge: Characterization of PFT properties

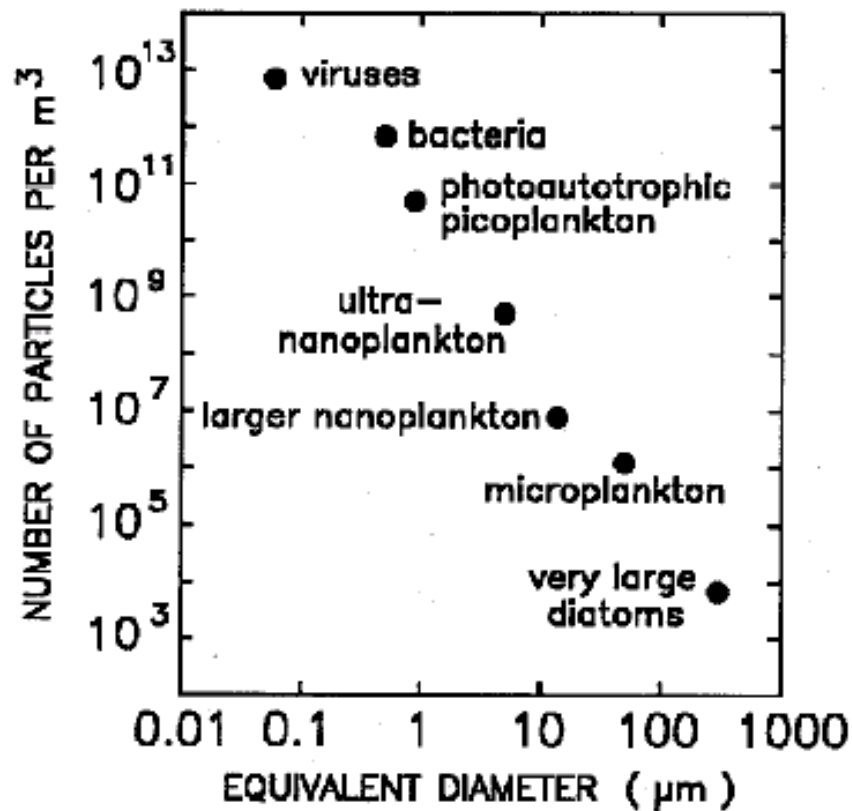
e.g., optical cross-sections $\sigma_i(\lambda)$

concentration in seawater N_i

Multi-component model of bulk IOPs

$$IOP(\lambda) = \sum_{i=1}^j IOP_i(\lambda) = \sum_{i=1}^j N_i \overline{\sigma}_i(\lambda)$$

$N_i \longrightarrow$

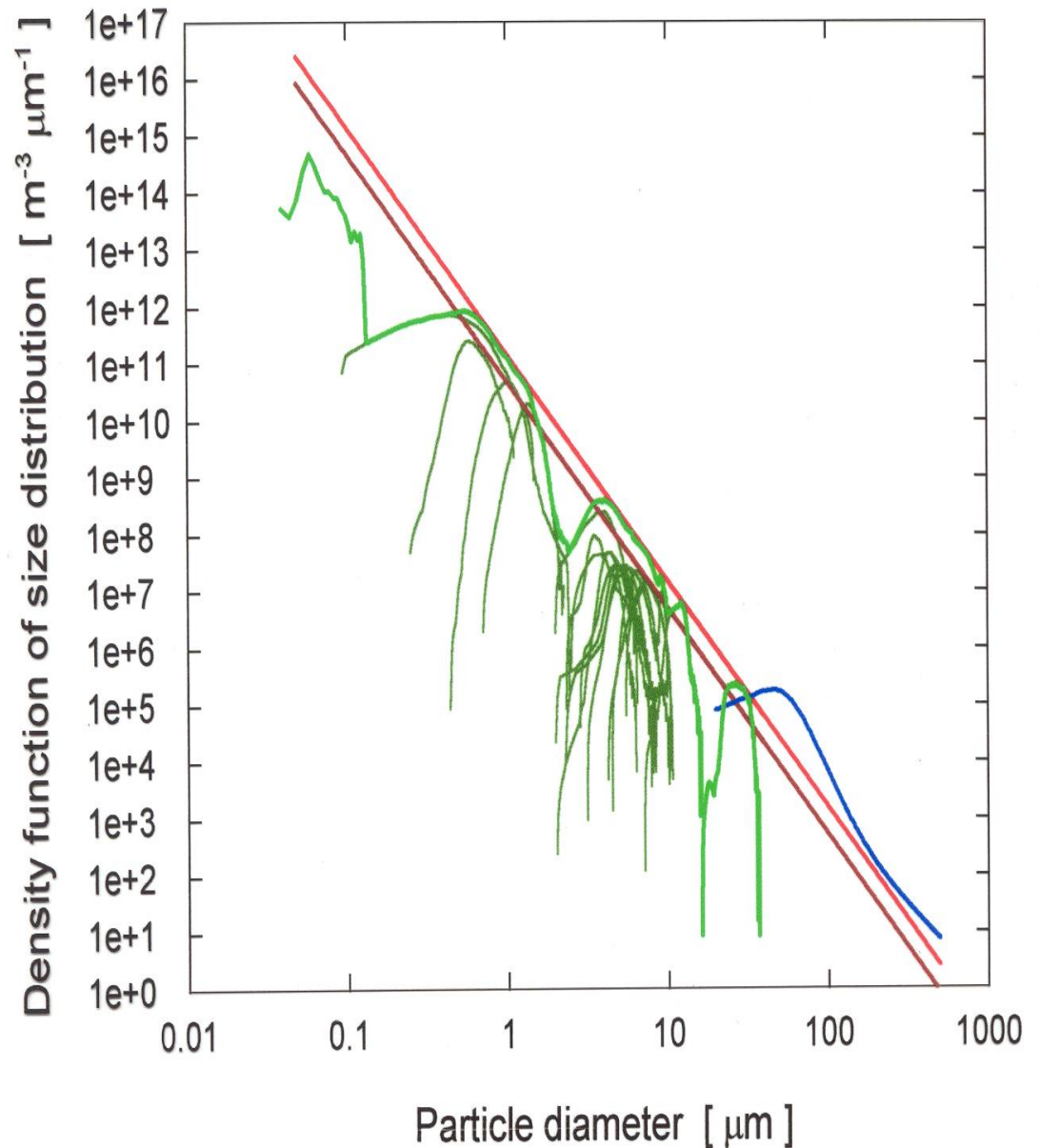


Example IOP model with detailed description of plankton community

i	Component	Concentration [particles/m ³]	Chl [mg m ⁻³]
1	VIRU	$1.0 \cdot 10^{13}$	0
2	HBAC	$4.0 \cdot 10^{11}$	0
3	PROC	$7.0 \cdot 10^{10}$	0.1026
4	SYNE	$2.0 \cdot 10^{10}$	0.0403
5	SYMA	$8.0 \cdot 10^9$	0.0360
Σ	Picoplankton	$4.98 \cdot 10^{11}$	0.1789
6	PING	$4.5056 \cdot 10^8$	0.0540
7	PSEU	$0.9808 \cdot 10^8$	0.0303
8	LUTH	$0.9924 \cdot 10^8$	0.0107
9	GALB	$0.4839 \cdot 10^8$	0.0155
10	HUXL	$0.4339 \cdot 10^8$	0.0104
11	CRUE	$0.4496 \cdot 10^8$	0.0129
12	FRAG	$0.4768 \cdot 10^8$	0.0157
13	PARV	$0.6247 \cdot 10^8$	0.0181
14	BIOC	$0.3966 \cdot 10^8$	0.0900
15	TERT	$0.3570 \cdot 10^8$	0.0609
16	CURV	$0.2987 \cdot 10^8$	0.0099
Σ	Small Nanoplankton	$1.0 \cdot 10^9$	0.3284
17	ELON	$1.7 \cdot 10^7$	0.1595
18	MICA	$2.0 \cdot 10^6$	0.0508
Σ	Total Plankton	$1.0499019 \cdot 10^{13}$	0.7176
19	DET	$3.3 \cdot 10^{14}$	0
20	MIN	$1.1 \cdot 10^{14}$	0
Σ	Total Non-living Particles	$4.4 \cdot 10^{14}$	0
21	BUB	$7.1 \cdot 10^6$	0

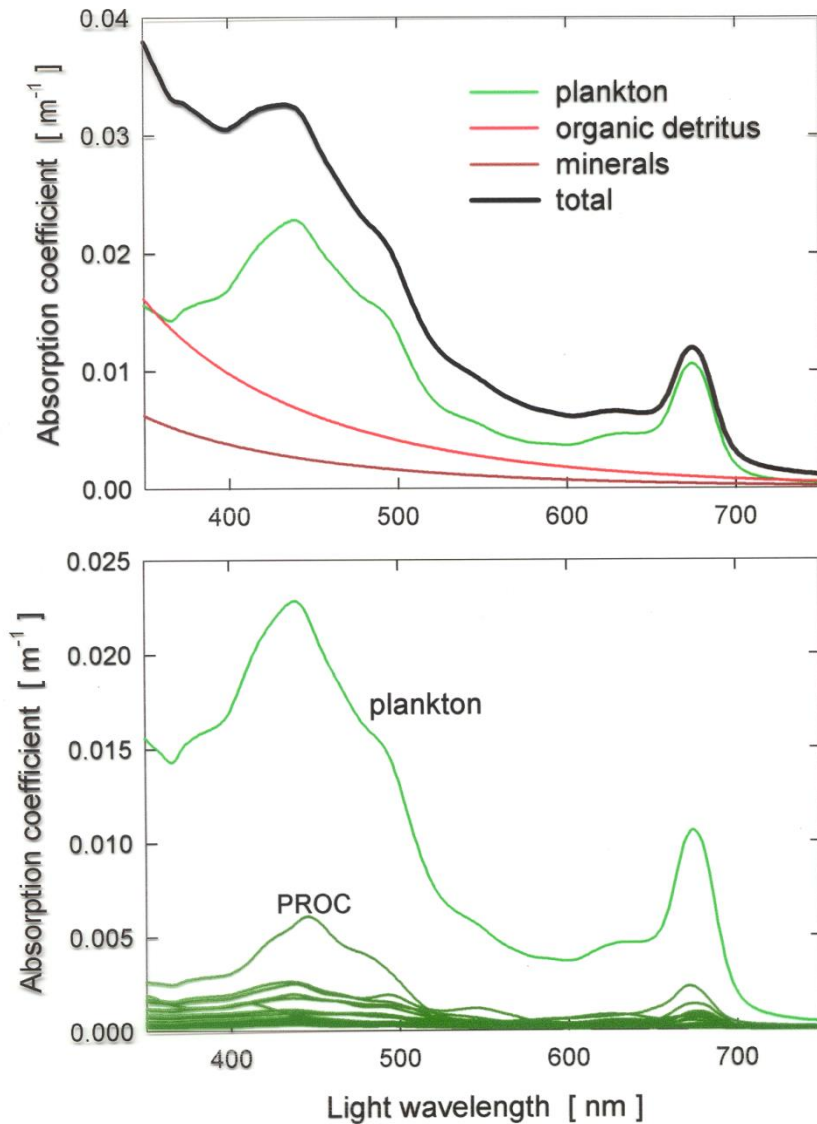
Size distribution

18 planktonic components
composite plankton
mineral particles
organic detritus
air bubbles

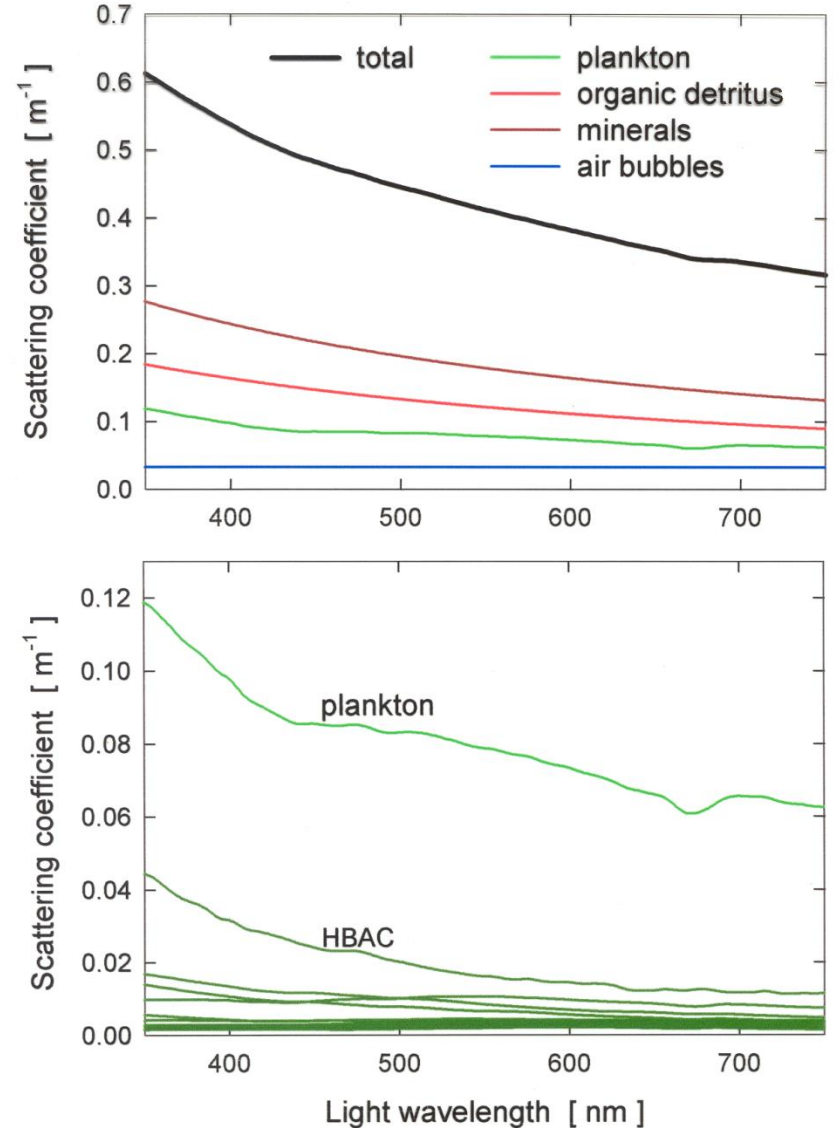


(Stramski et al. 2001)

Absorption



Scattering



Reductionist IOP/radiative transfer/reflectance model

Input to radiative transfer model

$$IOP(\lambda) = \sum_{i=1}^j IOP_i(\lambda) = \sum_{i=1}^j N_i \overline{\sigma}_i(\lambda)$$

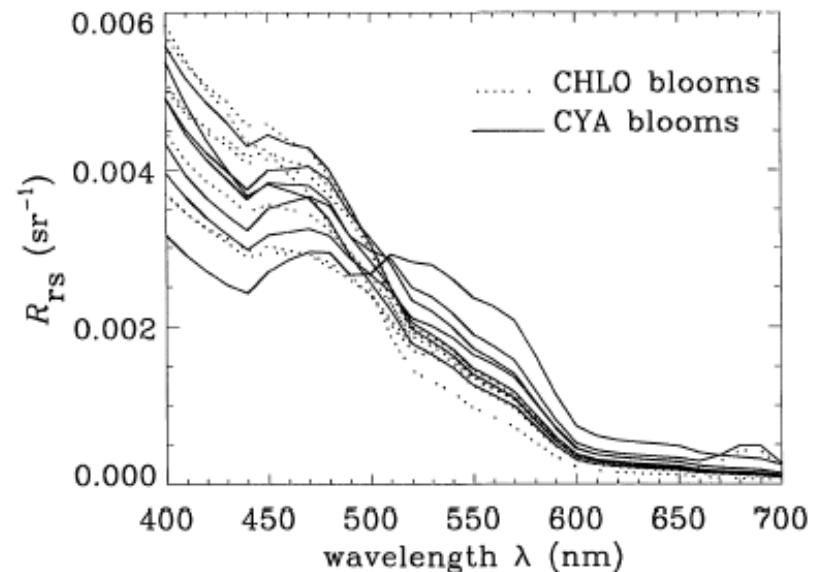
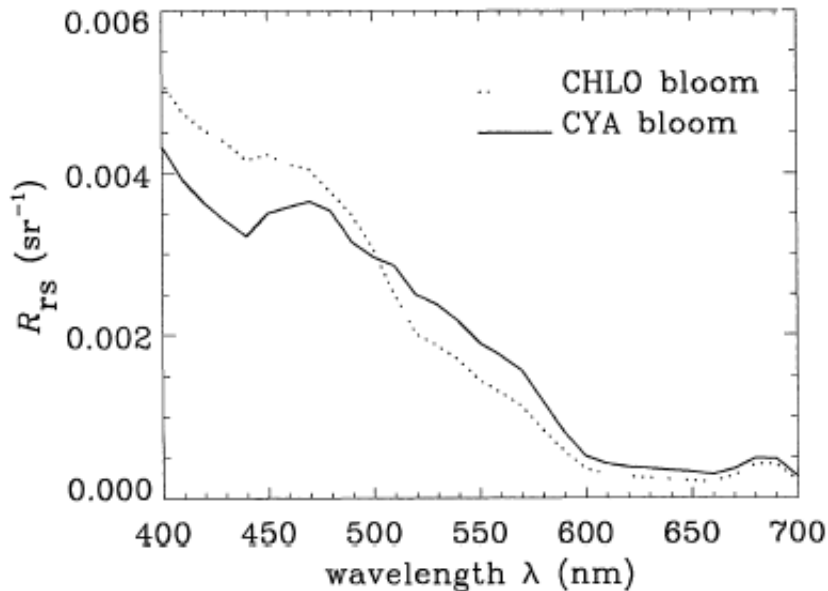
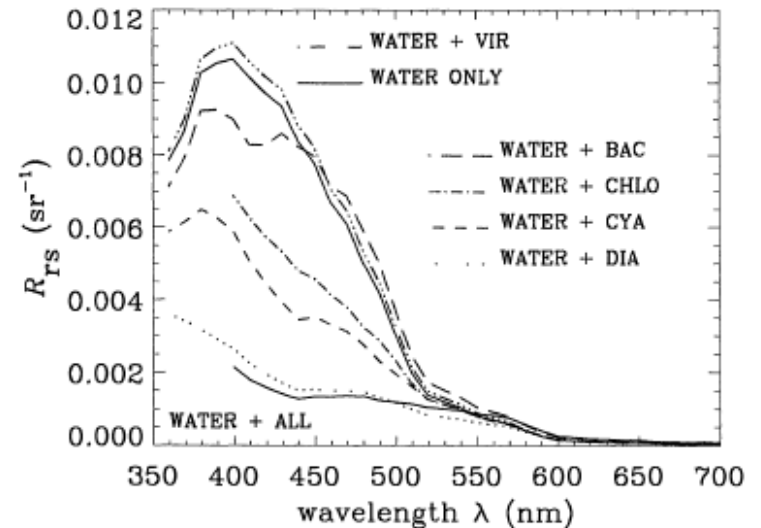
Output, e.g. ocean reflectance

$$R(\lambda) = f \left[\sum_{i=1}^j N_i \overline{\sigma}_{i,a}(\lambda), \sum_{i=1}^j N_i \overline{\sigma}_{i,b}(\psi, \lambda) \right]$$

- In what ways does variability in detailed seawater composition determine variability in ocean reflectance?
- What information about water constituents and optical properties can we hope to extract from remotely sensed reflectance?

Multi-component IOP database → Radiative transfer model

Viruses	$3 \times 10^9 - 1 \times 10^{14}$ particles m^{-3}
Heterotrophic bacteria	$1 \times 10^{11} - 2 \times 10^{12}$
Cyanobacteria	$1 \times 10^9 - 1 \times 10^{11}$ ($\text{Chl}^* = 4.63 \times 10^{-12}$)
Small diatoms	$3 \times 10^8 - 1 \times 10^{10}$ ($\text{Chl}^* = 3.09 \times 10^{-10}$)
Chlorophytes	$1 \times 10^7 - 4 \times 10^8$ ($\text{Chl}^* = 1.71 \times 10^{-9}$)
Detritus (DET)	$a_{\text{DET}}(400) = 0.2 - 0.6$ of $a_{\text{MICROBE}}(400)$ $b_{\text{DET}} = 0.2 - 0.6$ of $b_{\text{MICROBE}}(400)$
Yellow matter (CDOM)	$a_{\text{CDOM}}(400) = 0.2 - 0.4$ of $a_{\text{MICROBE}}(400)$ $b_{\text{CDOM}} = 0$



(Stramski and Mobley 1997; Mobley and Stramski 1997)

Reductionist approach: What do we need to do?

- Optical measurements
include $\beta(\psi, \lambda)$; target specific water constituents
- Particle identification and characterization
particle species composition, size distribution,
particle chemistry, biology, mineralogy, etc.
- Laboratory experiments (not just field experiments)
- New techniques and instrumentation

The complexity of seawater as an optical medium should not deter us from pursuing the proper course in future research

“The reductionist worldview has to be accepted as it is, not because we like it, but because that is the way the world works”

Steven Weinberg

1979 Nobel Prize in Physics